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Pneumococcal Surface Protein A: A Promising Candidate for the Next Generation of Pneumococcal Vaccines

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ABSTRACT

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Keywords: Pneumococcal vaccines; PspA; PspC; *S. pneumoniae* *Streptococcus pneumoniae* is the bacterium that causes pneumococcal disease which often results in pneumonia, meningitis, otitis media, septicemia and sinusitis. Pneumonia, particularly, is a significant cause of worldwide morbidity and a global health burden as well. Treatment often relies on antimicrobials, to which the pathogen is frequently mutating and rendering infective. Consequently, vaccination is the most effective approach in dealing with pneumococcal antimicrobial resistance (AMR). Unfortunately, the current pneumococcal polysaccharide and conjugate vaccines have a narrow serotype coverage. Therefore, the current need for vaccines with a broader serotype coverage cannot be overstated. Pneumococcal Surface Protein A and C are potential vaccine candidate antigens present in over 90% of the strains from clinical isolates as well as laboratory non-encapsulated strains. Pneumococcal Surface Protein A is an active virulent factor that pneumococci use to evade complement-mediated host immune responses and has been shown to elicit immune responses against pneumococcal infections. This review explores the potential utilization of Pneumococcal Surface Protein A to immunize against *S. pneumoniae*.

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Introduction

S. pneumoniae is a gram-positive bacterial pathogen that causes pneumococcal disease in humans. Infections of S. pneumoniae often result in pneumonia (the most common complication), meningitis, otitis media, septicemia, and sinusitis (1,2). Pneumococcal infections are considered a heavy global burden on the health systems, causing the highest number of deaths among infectious diseases (3). In normal individuals, the bacteria may be part of the normal human microbiome and reside asymptomatically in colonies in the upper respiratory tract and nasopharynx (4,5). However, any chance accorded to the pathogen, especially in immunocompromised persons, children and the elderly, results in invasive infections As of 2008, the Centers for Disease Control (CDC) projected approximately 1 million yearly deaths of children below the age of five resulting from pneumococcal infections, in the US alone (6).

Over a century of studying *S. pneumoniae* has led to a substantial understanding of the pathogen's physiology, pathogenesis, and immunity. Consequently, numerous antibiotics have been developed against the pathogen, and remain the mainstay of treatment in pneumococcal infections (7). The primary anti-pneumococcal drugs are β -lactams, macrolides and fluoroquinones (8). The tolerance of *S*.

*Corresponding author. E-mail: ssalghamdi@uqu.edu.sa; umar.sahibzada@gmail.com Cellular and Molecular Biology, 2021, 67(4): 289-298 pneumoniae to macrolides and β -lactams is a significant global concern. Research has shown that AMR in pneumococcus occurs in over 20% of the cases in some countries in Europe (9,10). The development of AMR in pneumococci has been linked to genetic transformation and selection of resistant pneumococci during asymptomatic carriage in children, who are often exposed to antibiotics. Therefore, restriction of fluoroquinolones' use in children explains why resistance rates against it remain relatively lower (11,12).

According to Castiglioni (2019), vaccination has long been regarded as the most impactful discovery in medicine from the public health perspective; however, it was not until recently that it was considered as the best solution to AMR. A publicly commissioned study in the UK dissected the topic deeply and reported the scientific economic benefits and of using immunization to fight AMR (13). There are two kinds of vaccines that are currently used to elicit immunity against S. pneumoniae: pneumococcal conjugate vaccines, and polysaccharide vaccines. PPSV23 is a 23-valent pneumococcal vaccine that protects against 23 capsular serotypes of the bacteria in individuals aged 2 years and above: however, it does not protect children below the age of 2. The 7-valent PCV7 was developed to protect against the 7 most common serotypes causing invasive pediatric pneumococcal disease. The most recent vaccine, PCV13, protects against 13 serotypes, in both adults and children (14-17). Daniels et al. (2016) proposed that the development of new and more virulent serotypes that are not protected by the vaccines reinforces the need to research into novel vaccine antigens (18).

The Pneumococcal Surface Protein A (PspA) is a cell-wall-associated antigen in *S. pneumonia*. It is a key virulent factor utilized by the pathogen to bind human lactoferrin and interfere with the opsonization of the bacteria (19,20). Roberts et al. (2019) have demonstrated that the protein offers broader protection against pneumococcal infections than the current pneumococcal conjugate vaccines (21). Yatim et al. (2013) observed PspA proteins in 95% of the strains affecting Malaysian children (22). Several other extensive studies have revealed over 90% consistency of the antigen among pneumococcal strains (23-25). Such consistency, combined with its role in the pathogenicity of the bacteria, renders PspA a

promising vaccine candidate. In this review, we explore the possible use of recombinant PspA as an antigen for immunization against *S. pneumoniae*.

Pneumococcal Virulence and Host Immunity

Pneumococci have a vast array of virulence factors (Fig. 1) that foster its attachment and invasion of host cells and allow it to evade the host's attempts to flush it out (26). A healthy and robust immune system is a prerequisite for flushing out pathogens before they are infective (27). Conversely, a weaker or incapable immune system provides a conduit for the attack even by the normally nonpathogenic microorganisms. One's immunity develops with age and gets stronger in adults, but it gets deteriorating in elderly people (28,29). S. pneumoniae exists asymptomatically in immunocompetent individuals, but those with weaker immune systems especially the children, elderly and immune-compromised persons have increased susceptibility to invasive infections resulting in pneumococcal disease (30,31).

The capacity of pneumococci to expand their genetic material through transformation and recombination is the primary technique for their virulence (32). The degree of genetic diversity within S. pneumoniae must be investigated in order to fully comprehend its pathogenicity as well as create viable therapies and vaccines (33). The virulence profile of pneumococcal strains is several optimized by variations in genetic content and single genes. Genome diversification was defined by Zhao et al. (2018) as bacterial ability to develop in a variety of host settings (34). Because of variations in the genetic content of their dispensable genes, genetic diversity has been found among identical clones of the pathogen. Dispensable genes aren't required for bacterial growth; rather they give the pathogen selection benefits like antibiotic resistance. Allele replacement introduces additional variations to the microbe's core genome. This is due to the bacteria's absence of SOS genes, which prevent it from repairing damaged DNA. Genetic variation can also be influenced by pregnancy. Chaguza et al., (2020) established a model to estimate carriage time and combined their results with whole-genome sequencing (WGS) data. In comparison to the patient's age and prior carriage, the WGS data showed that bacterial

genetic variation accounted for phenotypic variance (35).

Critical virulence factors in *S. pneumoniae* infections include: capsular and cell wall polysaccharides, choline-binding proteins (CBPs), Pneumococcal Surface Protein A (PspA), Pneumococcal Surface Protein C (PspC), pneumolysin, autolysin, among others.

Immune responses mounted against pneumococcal infections are mediated through both arms of the immune system: innate and adaptive (36) Innate immunity is mediated through the mucosa and respiratory epithelial cells, phagocytic cells and pattern recognition receptors (17, 37, 38). Adaptive immune responses, which are elicited a few days after the infection, are mediated through B and T Pneumococcal-specific lymphocytes (39). IgA antibody, secreted at infection sites, is essential for opsonizing the S. pneumoniae pathogens and promoting phagocytosis (40). IgA1 protease possessed by the pathogen, however, cleaves IgA and prevents opsonization. The binding of the Fab fragment to the cell wall following IgA cleavage exposes CBPs, lowers the capsular negative charge and causes the bacterial cells to attach more firmly (41-43). Simultaneous activation stimulates differentiation of naïve B cells into IgM-positive memory B cells and facilitates class switching to produce other Ig types required to flush out the infection (44).

Presentation of antigen peptide-MHC complexes by APCs stimulates T cell response. CD4 T helper cells are activated by co-stimulatory proteins and initiate a cascade of events geared towards generating both cellmediated and humoral responses against the infection (45,46). Immune responses following pneumococcal infections hampered are often in immunocompromised individuals. Infant T cell responses fail because exposure to foreign antigens is often restricted before birth. The efficacy of adaptive responses also diminishes in aged individuals and they are thus susceptible to morbidity (47).

As already established, capsular polysaccharides and pneumolysin are among the primary pneumococcal virulence factors, and existing vaccines are based on these two (48). However, a few studies have described the existence of non-encapsulated pathogenic strains; and these are usually unaffected by the existing vaccines (49-51). This, alongside the discovery of new virulence factors and new methods of pathogenesis for existing virulence factors, demonstrates the pneumococcus' strength in the face of environmental obstacles, particularly those presented by antimicrobials and vaccines. Recent advancements in our grasp of pneumococcal virulence factors may open the door to the creation of innovative treatment or preventative methods.

Novel vaccine approaches

The main antigenic constituents of the present pneumococcal vaccines are capsular polysaccharides. However, polysaccharide-based vaccines are unable to protect children aged 2 years and below. Conjugate vaccines have therefore been developed by coupling immunogenic proteins with the capsules. These vaccines elicit responses efficiently, but only against a limited number of serotypes. The most recent development approaches, therefore, include using immunogenic proteins that are consistent in a wide array of serotypes (52).

Immunogenic proteins of *S. pneumoniae* that contribute to its virulence are an avenue for developing vaccines with a wider serotype coverage. Some of the proteins include PspA, PspC, PsaA, PppA, Zinc metalloprotease B, among others (53,54). Table 1 illustrates some of the pneumococcal vaccines that are currently in use and those that are still in clinical trials. PspA and PspC have been widely studied as critical pneumococcal virulence factors expressed by virtually all strains, and exhibit diverse organ-specific effects (55-57).



Figure 1. Common pneumococcal virulence factors (36).

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S	Pneumococcal Vaccine	Туре	Status
No.	Candidates		
1	23-Valent Pneumococcal	Polysaccharide	А
	Polysaccharide Vaccine		
	(PPV23)		
2	Pneumococcal Conjugate	Conjugate	А
	Vaccine		
	(PCV-7, PCV-10 and PCV-		
	13)		
3	Pneumococcal Conjugate	Conjugate	В
	Vaccine		
	(PCV-15 and PCV-20)		
4	Pneumococcal Protein	Protein	В
	Vaccine		
	(PspA, PhtD, SktP and		
	-		
	pneumolysin)		

Table 1. Current pneumococcal vaccines in use and vaccine candidates in clinical trials; in use (A) and in clinical trials (B)

Pneumococcal Surface Protein A

PspA is a CBP protein that inhibits cell surface binding of C3 complement constituent thus interfering with opsonization (58). The protein is present in clinical isolates, as well as non-encapsulated strain Rx1. PspA was identified with the help of monoclonal antibodies that could elicit pneumococcal immune responses in mice (59). Immunizing CBA/N mice with congenic PspA+ and PspA- pathogens has proven that PspA elicits protective anti-pneumococcal antibodies (60). Insertional inactivation of the gene that codes for PspA in three S. pneumoniae strains has also been shown to reduce the virulence of all three, with two becoming completely avirulent (61,62). These observations are significant to future pneumococcal vaccine R&D, as CBA/D mice, like infants and elderly persons, are unresponsive to the current pneumococcal vaccines (63,64).

Following an extensive experimental study, Tu et al. (1999) reported that in mice infected with *S. pneumoniae*, PspA inhibits complement-dependent host immune responses mediated by factor B (65). Immunoblots of opsonized bacterial cells showed that C3b was present on PspA- bacteria, but not on PspA+ bacteria. In addition, the α -chain of C3b was cleaved and its processing was lowered by PspA. It can therefore be inferred that the virulence of PspA is by blocking the opsonic binding of C3b on the pathogen's surface or by inhibiting the functionality of the alternative pathway's C3 convertase. When the later mechanism is utilized, PspA lowers the quantity

of C3b deposited on the bacterial surfaces, effectively inhibiting complement-mediated pathways of bacterial clearance (44,57,66).

Being a highly variable protein, PspA can be grouped into three families and six different clades based on the sequence of its amino end (67). The protein elicits immune responses against fatal septicemia resulting from different pneumococcal serotypes. The N-terminal of PspA comprises repetitive helices protruding from the cell surface. The region between the two terminals of the protein, consisting of 60-80 amino acids, is particularly rich in proline, and also highly variable in both length and sequence. Antibodies targeted against this portion of the protein are cross-reactive but not cross-protective (68,69).

Immunization with PspA

PspA is In mice models, the strongest immune response is elicited when the immunizing PspAs belong to a similar family as the challenge PspAs (70). This emphasizes the need to include proteins from different clades and families when developing PspA-based vaccines for protection in humans. Administering both PspA and IL-12 intranasal has been demonstrated to increase the secretion of systemic antibodies, enhance opsonization, and confer stronger protection (71,72). Consequently, IL-12 could be an effective adjuvant for PspA-based immunization. Also, a toll-like receptor 5 binding protein found in Vibrio vulnificus, FlaB, exhibits a high mucosal adjuvant activity. Experimental studies in mice have shown the potential role of FlaB in PspA immunization. In these experiments, one group of mice was immunized with recombinant fusion proteins made of both PspA and FlaB, the second group was immunized with a direct mixture of PspA and FlaB, while the control group was immunized with PspA alone. The findings revealed more protection in groups one and two than in the control group. However, group one elicited a much stronger immune response than the second group, we can therefore infer from these studies that genetic recombination of FlaB and PspA is necessary for the high efficacy of FlaB-adjuvanted S. pneumoniae vaccines (73).

Phase 1 clinical trials of a recombinant PspA vaccine belonging to family 1 have been completed in

man, and the vaccine is safe and immunogenic. In addition, when the immune human serum was administered to mice infected with pneumococcal pathogens expressing either PspA family 1 or 2, an immune response was elicited, but not as strong against family 2 as against family 1. This reinforces the need for using IL-12 or FlaB-adjuvanted recombinant pneumococcal vaccines (74).

PspC, also known as CbpA, is another vaccine candidate that has been extensively studied (75). PspC binds secretory IgA to enable it to bind the host's epithelial and endothelial cells. Immunizing mice with PspC protects against pneumococcal septicemia. Moreover, Daniels et al. (2010) showed that antibodies targeted against PspC exhibit crossreactivity against PspA (76). This cross-reactivity has been attributed to the many similarities that exist between the two surface proteins (77). Both proteins have a proline-rich region in the α -helix between the N and the C terminals. Within the proline-rich regions of the two proteins, there exists an invariable nonproline block (NPB) with a sequence of thirty-three amino acids. Therefore, immunization with recombinant proline-rich molecules and monoclonal antibodies against the NPB or proline-rich epitopes elicits a robust immune response in mice (78,79). Because the proline-rich and NPB regions are highly conserved ad cross-reactive, they represent potential targets for PspA and PspC-based recombinant vaccines. Several studies have examined both PspA and PspC as active pneumococcal virulence factors, present consistently in over 90% of all S. pneumoniae strains, and a majority of them have demonstrated the importance of combining the two (62,80-82). Schachern et al. (2014) did extensive research on the immunization capabilities of PspA, PspC, and both PspA and PspC. From their findings, they were able to demonstrate that PspC has the ability to dramatically boost the efficacy of a multi-component PspA vaccine (83,84).

Conclusions

PspA is a critical virulence factor in pneumococci that acts by blocking complement-mediated opsonization of the pathogens. This prevents phagocytosis, effectively inhibiting clearance of the pneumococci. PspA has a greater capacity to elicit stronger immune responses with a broad serotype coverage than the current polysaccharide-based and pneumococcal conjugate vaccines. Polysaccharidebased vaccines do not offer protection to infants, while both of them have only limited serotype coverage and also offer zero protection against nonencapsulated strains. PspA antigens are present in over 90% of the pneumococcal strains, including nonencapsulated ones. Immunization targeting this antigen provides a wider serotype coverage and is thus more efficient. Pneumococcal vaccine R&D should employ a recombinant approach that integrates both PspA and PspC in a multi-component vaccine, because of their combined stronger immune response and even wider serotype coverage resulting from cross-reactivity. A combination of all the three families of the PspA antigen in the same vaccine is also recommended, to eliminate the negative effects of the phenotypic variations that could result from the existence of different PspA families. IL-12 and FlaB are efficient adjuvants that can be utilized to enhance the performance of PspA-based vaccines. However, FlaB-adjuvanted PspA vaccines would require genetic recombination of protein segments from both PspA and FlaB, rather than a mere mixture of the two, to function efficiently.

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References

- de Gier B, van Kassel MN, Sanders EA, van de Beek D, Hahné SJ, van der Ende A, et al. Disease burden of neonatal invasive Group B Streptococcus infection in the Netherlands. PloS one 2019; 14(5): e0216749.
- 2. Jacobs MR. *Streptococcus pneumoniae*: epidemiology and patterns of resistance. The Am J M Suppl 2004; 117(3): 3-15.
- Henriques-Normark B, Tuomanen EI. The pneumococcus: epidemiology, microbiology, and pathogenesis. Cold Spring Harb Perspect Med 2013; 3(7): a010215.
- 4. Ahmadi A, Yaghoubi S, Irajian G. Molecular analysis of PBP1A in *Streptococcus pneumoniae* isolated from clinical and normal Flora samples in Tehran, Iran: a multicenter study. Microb Drug Resist 2019; 25(1): 39-46.

- DeMuri GP, Gern JE, Eickhoff JC, Lynch SV, Wald ER. Dynamics of bacterial colonization with *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Moraxella catarrhalis* during symptomatic and asymptomatic viral upper respiratory tract infection. Clin Infect Dis 2018; 66(7): 1045-1053.
- Centers for Disease Control and Prevention (CDC). Progress in introduction of pneumococcal conjugate vaccine--worldwide, 2000-2008. MMWR. Morb Mortal Wkly Rep, 2008; 57(42): 1148-1151.
- Magill SS, O'Leary E, Janelle SJ, Thompson DL, Dumyati G, Nadle J, et al. Changes in prevalence of health care–associated infections in US hospitals. N Engl J Med 2018; 379(18): 1732-44.
- Cillóniz C, Garcia-Vidal C, Ceccato A, Torres A. Antimicrobial resistance among *Streptococcus pneumoniae*. In Antimicrobial Resistance in the 21st Century 2018;13-38. Springer, Cham.
- Linares J, Ardanuy C, Pallares R, Fenoll, A. Changes in antimicrobial resistance, serotypes and genotypes in *Streptococcus pneumoniae* over a 30-year period. Clin Microbiol infect 2010; 16(5): 402-410.
- Lynch JP, & Zhanel GG. Streptococcus pneumoniae: does antimicrobial resistance matter? In Seminars in Respiratory and Critical Care Medicine 2009; 30(02): 210-238. © Thieme Medical Publishers.
- Chen HH, Stringer A, Eguale T, Rao GG, Ozawa S. Impact of antibiotic resistance on treatment of pneumococcal disease in Ethiopia: an agent-based modeling simulation. Am J Trop Med Hyg 2019; 101(5): 1042.
- Saleem Z, Hassali MA, Godman B, Versporten A, Hashmi FK, Saeed H. Point prevalence surveys of antimicrobial use: a systematic review and the implications. Expert Rev Anti Infect Ther 2020; 18(9): 897-910.
- 13. O'Neil J. Tackling Drug-Resistant Infections Globally. Rev Antimicrob Resist. 2016.
- Colijn C, Corander J, Croucher NJ. Designing ecologically optimized pneumococcal vaccines using population genomics. Nat microbiol 2020; 5(3): 473-485.

- 15. Feldman C, Anderson R. Current and new generation pneumococcal vaccines. J Infect 2014; 69(4): 309-325.
- 16. Hurley LP, Allison MA, Pilishvili T, O'Leary ST, Crane LA, Brtnikova M, et al. Primary care physicians' struggle with current adult pneumococcal vaccine recommendations. J Am Board Fam Med 2018; 31(1): 94-104.
- 17. Alghamdi S. The Role of Vaccines in Combating Antimicrobial Resistance (AMR) Bacteria. Saudi J Biol Sci 2021.
- Daniels CC, Rogers PD, Shelton CM. A review of pneumococcal vaccines: current polysaccharide vaccine recommendations and future protein antigens. J Pediatr Pharmacol Ther 2016; 21(1): 27-35.
- 19. Mukerji R, Hendrickson C, Genschmer KR, Park SS, Bouchet V, Goldstein R, et al. The diversity of the proline-rich domain of pneumococcal surface protein A (PspA): Potential relevance to a broad-spectrum vaccine. Vaccine 2018 36(45), 6834-6843.
- 20. Park SS, Gonzalez-Juarbe N, Martínez E, Hale JY, Lin YH, Huffines JT, et al. *Streptococcus pneumoniae* binds to host lactate dehydrogenase via PspA and PspC to enhance virulence. Mbio 2021; 12(3): e00673-21.
- 21. Roberts S, Williams CM, Salmon SL, Bonin JL, Metzger DW, & Furuya Y. Evaluation of pneumococcal surface protein A as a vaccine antigen against secondary *Streptococcus pneumoniae* challenge during influenza A infection. Vaccines 2019; 7(4): 146.
- 22. Yatim MM, Masri SN, Desa MNM, Taib NM, Nordin SA, Jamal F. Determination of phenotypes and pneumococcal surface protein A family types of *Streptococcus pneumoniae* from Malaysian healthy children. Journal of Microbiology, Immunol Infect 2013; 46(3): 180-186.
- Beall B, Gherardi G, Facklam RR, Hollingshead SK. Pneumococcal pspA sequence types of prevalent multiresistant pneumococcal strains in the United States and of internationally disseminated clones. J Clin Microbiol 2000; 38(10): 3663-3669.
- 24. Kothari N, Kothari S, Choi YJ, Dey A, Briles DE, Rhee DK, et al. A bivalent conjugate vaccine containing PspA families 1 and 2 has the potential

to protect against a wide range of *Streptococcus pneumoniae* strains and *Salmonella Typhi*. Vaccine 2015; 33(6): 783-788.

- 25. Morino S, Kitagami E, Nakayama H, Koizumi Y, Tanaka-Taya Kinjo Y. K, et al. Seroepidemiological analysis of antipneumococcal surface protein А (PspA) immunoglobulin G by clades in Japanese population. Vaccine 2020; 38(47): 7479-7484.
- 26. Brooks LR, Mias GI. *Streptococcus pneumoniae's* virulence and host immunity: aging, diagnostics, and prevention. Front Immunol 2018; 9, 1366.
- Frank SA. Immunology and evolution of infectious disease. Princeton University Press. 2020
- 28. Burggraf V. Healthy People 2020: Implications for Practice. Healthy Aging 2020; 9.
- 29. Alghamdi S. Isolation and identification of the oral bacteria and their characterization for bacteriocin production in the oral cavity. Saudi J Biol Sci 2021.
- 30. Lejri-El Euchi H, Chirpaz E, Foucher A, Sultan-Bichat N, Randrianjohany A, Poubeau P, et al. Vaccination against influenza and pneumococcal infections in patients with autoimmune disorders under biological therapy: coverage and attitudes in patients and physicians. Eur J Intern Med 2019; 69, 25-31.
- 31. Renko M, Kukkola HL, Kauma H, Tapiainen T, Kaijalainen T, Uhari M. Comparison of the severity and outcome of invasive pneumococcal infections in children and adults. Pediatr Infect Dis J 2012; 31(7): 785-788.
- Kilian M, Tettelin H. Identification of virulenceassociated properties by comparative genome analysis of *S. pneumoniae*, *S. pseudopneumoniae*, *S. mitis*, three *S. oralis* subspecies, and *S.* infantis. MBio 2019; 10(5): e01985-19.
- Tettelin H, Chancey S, Mitchell T, Denapaite D, Schähle Y, Rieger M. Genomics, genetic variation, and regions of differences. In *Streptococcus pneumoniae* 2015; 81-107. Academic Press.
- 34. Zhao Y, Sun C, Zhao D, Zhang Y, You Y, Jia X, et al. PGAP-X: extension on pan-genome analysis pipeline. BMC Genom 2018; 19(1): 115-124.
- 35. Chaguza C, Yang M, Cornick JE, Du Plessis M, Gladstone RA, Kwambana-Adams BA, et al.

Bacterial genome-wide association study of hyper-virulent pneumococcal serotype 1 identifies genetic variation associated with neurotropism. Commun Biol 2020; 3(1): 1-12.

- 36. Brooks LR, Mias GI. *Streptococcus pneumoniae's* virulence and host immunity: aging, diagnostics, and prevention. Front Immunol 2018; 9, 1366.
- 37. Lee HJ, Woo Y, Hahn TW, Jung YM, Jung YJ. Formation and maturation of the phagosome: a key mechanism in innate immunity against intracellular bacterial infection. Microorganisms 2020; 8(9): 1298.
- 38. Weight CM, Venturini C, Pojar S, Jochems SP, Reiné J, Nikolaou E, et al. Microinvasion by *Streptococcus pneumoniae* induces epithelial innate immunity during colonisation at the human mucosal surface. Nat Commun 2019; 10(1): 1-15.
- 39. Koh SH, Shin SG, Andrade MJ, Go RH, Park S, Woo CH, et al. Long pentraxin PTX3 mediates acute inflammatory responses against pneumococcal infection. Biochem Biophys Res Commun 2017; 493(1): 671-676.
- 40. Riegler AN, Brissac T, Gonzalez-Juarbe N, Orihuela CJ. Necroptotic cell death promotes adaptive immunity against colonizing pneumococci. Front Immunol 2019; 10: 615.
- 41. Chi YC, Rahkola JT, Kendrick AA, Holliday MJ, Paukovich N, Roberts TS, et al. *Streptococcus pneumoniae* IgA1 protease: A metalloprotease that can catalyze in a split manner in vitro. Protein Sci 2017; 26(3): 600-610.
- 42. Marquart ME. Pathogenicity and virulence of *Streptococcus pneumoniae*: Cutting to the chase on proteases. Virulence 2021; 12(1): 766-787.
- Wang Z, Rahkola J, Redzic JS, Chi YC, Tran N, Holyoak T, et al. Mechanism and inhibition of *Streptococcus pneumoniae* IgA1 protease. Nat Commun 2020; 11(1): 1-8.
- 44. Andre GO, Converso TR, Politano WR, Ferraz LF, Ribeiro ML, Leite LC, et al. Role of *Streptococcus pneumoniae* proteins in evasion of complement-mediated immunity. Front Microbiol 2017; 8: 224.
- 45. Kurioka A, Van Wilgenburg B, Javan RR, Hoyle R, Van Tonder AJ, Harrold CL, et al. Diverse *Streptococcus pneumoniae* Strains Drive a Mucosal-Associated Invariant T-Cell Response Through Major Histocompatibility Complex class

I–Related Molecule–Dependent and Cytokine-Driven Pathways. J Infect Dis 2018; 217(6): 988-999.

- 46. Middleton DR, Sun L, Paschall AV, Avci FY. T cell-mediated humoral immune responses to type 3 capsular polysaccharide of *Streptococcus pneumoniae*. J Immunol 2017; 199(2): 598-603.
- 47. Smith EL, Adler H, Ferreira DM, Sá-Leão R, Abdullahi O, Adetif I, et al. Upper airways colonisation of *Streptococcus pneumoniae* in adults aged 60 years and older: a systematic review of prevalence and individual participant data meta-analysis of risk factors. J Infect 2020; 81(4): 540-548.
- 48. Hupp S, Grandgirard D, Mitchell TJ, Leib SL, Hathaway LJ, Iliev AI. Pneumolysin and the bacterial capsule of *Streptococcus pneumoniae* cooperatively inhibit taxis and motility of microglia. J Neuroinflammation 2019; 16(1): 1-14.
- 49. Bradshaw JL, Pipkins HR, Keller LE, Pendarvis JK, McDaniel LS. Mucosal infections and invasive potential of nonencapsulated *Streptococcus pneumoniae* are enhanced by oligopeptide binding proteins AliC and AliD. MBio 2018; 9(1): e02097-17.
- 50. Bradshaw JL, Rafiqullah IM, Robinson DA, McDaniel LS. Transformation of nonencapsulated *Streptococcus pneumoniae* during systemic infection. Sci Rep 2020; 10(1): 1-9.
- 51. Takeuchi N, Ohkusu M, Wada N, Kurosawa S, Miyabe A, Yamaguchi M, et al. Molecular typing, antibiotic susceptibility, and biofilm production in nonencapsulated *Streptococcus pneumoniae* isolated from children in Japan. J Infect Chemother 2019; 25(10): 750-757.
- 52. Croucher NJ, Løchen A, Bentley SD. Pneumococcal vaccines: host interactions, population dynamics, and design principles. Annu Rev Microbiol 2018; 72: 521-549.
- 53. Lagousi T, Basdeki P, De Jonge MI, Spoulou V. Understanding host immune responses to pneumococcal proteins in the upper respiratory tract to develop serotype-independent pneumococcal vaccines. Expert Rev Vaccines 2020; 19(10): 959-972.
- 54. Lagousi T, Basdeki P, Routsias J, Spoulou V. Novel protein-based pneumococcal vaccines:

assessing the use of distinct protein fragments instead of full-length proteins as vaccine antigens. Vaccines 2019; 7(1): 9.

- 55. Azarian T, Grant LR, Georgieva M, Hammitt LL, Reid R, Bentley SD, et al. Association of pneumococcal protein antigen serology with age and antigenic profile of colonizing isolates. J infect Dis 2017; *215*(5): 713-722.
- 56. Du S, Vilhena C, King S, Sahagún-Ruiz A, Hammerschmidt S, Skerka C, et al. Molecular analyses identifies new domains and structural differences among *Streptococcus pneumoniae* immune evasion proteins PspC and Hic. Sci Rep 2021; 11(1): 1-15.
- 57. Haleem KS, Ali YM, Yesilkaya H, Kohler T, Hammerschmidt S, Andrew PW, et al. The pneumococcal surface proteins PspA and PspC sequester host C4-binding protein to inactivate complement C4b on the bacterial surface. Infect Immun, 2018; 87(1): e00742-18.
- 58. Rodrigues TC, Oliveira MLS, Soares-Schanoski A, Chavez-Rico SL, Figueiredo DB, Gonçalves VM, et al. Mucosal immunization with PspA (Pneumococcal surface protein A)-adsorbed nanoparticles targeting the lungs for protection against pneumococcal infection. PloS one 2018; 13(1): e0191692.
- 59. Briles DE, Yother J, McDaniel LS. Role of pneumococcal surface protein A in the virulence of *Streptococcus pneumoniae*. Rev Infect Dis 1988; S372-S374.
- 60. Wu HY, Nahm MH, Guo Y, Russell MW, Briles DE. Intranasal immunization of mice with PspA (pneumococcal surface protein A) can prevent intranasal carriage, pulmonary infection, and sepsis with *Streptococcus pneumoniae*. J Infect Dis 1997; 175(4): 839-846.
- 61. McDANIEL LS, Yother JANET, Vijayakumar M, McGARRY LYNN, Guild WR, Briles, DE. Use of insertional inactivation to facilitate studies of biological properties of pneumococcal surface protein A (PspA). J Exp Med 1987;165(2): 381-394.
- 62. Park SS, Gonzalez-Juarbe N, Martínez E, Hale JY, Lin YH, Huffines JT, et al. *Streptococcus pneumoniae* binds to host lactate dehydrogenase via PspA and PspC to enhance virulence. Mbio 2021; 12(3): e00673-21.

- 63. Huang J, Gingerich AD, Royer F, Paschall AV, Pena-Briseno A, Avci FY, Mousa JJ. Broadly reactive human monoclonal antibodies targeting the pneumococcal histidine triad protein protect against fatal pneumococcal infection. Infect Immun 2021; 89(5): e00747-20.
- 64. Medina M, Villena J, Vintiñi E, Hebert EM, Raya R, Alvarez S. Nasal immunization with Lactococcus lactis expressing the pneumococcal protective protein A induces protective immunity in mice. Infect Immun 2008; 76(6): 2696-2705.
- 65. Tu AHT, Fulgham RL, McCrory MA, Briles DE, Szalai AJ. Pneumococcal surface protein A inhibits complement activation by *Streptococcus pneumoniae*. Infect Immun 1999; 67(9): 4720-4724.
- 66. Yuste J, Botto M, Paton JC, Holden DW, Brown JS. Additive inhibition of complement deposition by pneumolysin and PspA facilitates *Streptococcus pneumoniae* septicemia. J Immunol 2005; 175(3): 1813-1819.
- 67. Yu J, Chen X, Li B, Gu T, Meng X, Kong W, Wu Y. A pneumococcal vaccine combination with two proteins containing PspA families 1 and 2 can potentially protect against a wide range of *Streptococcus pneumoniae* strains. Immunol Res 2018; 66(4): 528-536.
- 68. Converso TR, Goulart C, Rodriguez D, Darrieux M, Leite LCC. Rational selection of broadly cross-reactive family 2 PspA molecules for inclusion in chimeric pneumococcal vaccines. Microb Pathog 2017; 109: 233-238.
- 69. Knupp-Pereira PA, Marques NTC, Teixeira LM, Póvoa HCC, Neves FPG. Prevalence of PspA families and pilus islets among *Streptococcus pneumoniae* colonizing children before and after universal use of pneumococcal conjugate vaccines in Brazil. Brazilian J Microbiol 2020; *51*(2): 419-425.
- 70. Liu N, Dong Z, Zhu X, Xu H, Zhao Z. Characterization and protective effect of Polygonatum sibiricum polysaccharide against cyclophosphamide-induced immunosuppression in Balb/c mice. Int J Biol Macromol 2018; 107: 796-802.
- Aljewari H, de Castro R, Solomon O, Moore III QC, Nave F, Thompson A. Study of Pneumococcal Surface Protein, PspA,

Incorporated in Poly (Vinyl Alcohol) Hydrogel Membranes. J Biomater Nanobiotechnol 2020; 11(01): 67.

- 72. Goulart C, Rodriguez D, Kanno AI, Lu YJ, Malley R, Leite LC. Recombinant BCG expressing a PspA-PdT fusion protein protects mice against pneumococcal lethal challenge in a prime-boost strategy. Vaccine 2017; 35(13): 1683-1691.
- 73. Walkowski W, Bassett J, Bhalla M, Pfeifer BA, Ghanem ENB. Intranasal Vaccine Delivery Technology for Respiratory Tract Disease Application with a Special Emphasis on Pneumococcal Disease. Vaccines 2021; 9(6): 589.
- 74. Akbari E, Negahdari B, Faraji F, Behdani M, Kazemi-Lomedasht F, Habibi-Anbouhi M. Protective responses of an engineered PspA recombinant antigen against *Streptococcus pneumoniae*. Biotechnol Rep 2019; 24: e00385.
- 75. Tarahomjoo S. Recent approaches in vaccine development against *Streptococcus pneumoniae*. J Mol Microbiol Biotechnol 2014; 24(4): 215-227.
- 76. Daniels CC, Coan P, King J, Hale J, Benton KA, Briles DE, et al. The proline-rich region of pneumococcal surface proteins A and C contains surface-accessible epitopes common to all pneumococci and elicits antibody-mediated protection against sepsis. Infect Immun 2010; 78(5): 2163-2172.
- 77. Shahriar M. Antigenic cross reactivity between the pneumococcal polysaccharide vaccine and the *streptococcus pneumoniae* 7f one of the prevalent sertypes in Bangladesh (Doctoral dissertation, BRAC University). 2008.
- 78. Houben T, Magro dos Reis I, Oligschlaeger Y, Steinbusch H, Gijbels MJ, Hendrikx T,et al. Pneumococcal immunization reduces neurological and hepatic symptoms in a mouse model for Niemann-Pick type C1 disease. Front Immunol 2019; 9: 3089.
- 79. Ogunniyi AD, Folland RL, Briles DE, Hollingshead SK, Paton JC. Immunization of mice with combinations of pneumococcal virulence proteins elicits enhanced protection against challenge with *Streptococcus pneumoniae*. Infection and immunity 2000; 68(5): 3028-3033.

- Jedrzejas MJ. Pneumococcal virulence factors: structure and function. Microbiol Mol Biol Rev 2001; 65(2): 187-207.
- 81. Kadioglu A, Weiser JN, Paton JC, Andrew PW. The role of *Streptococcus pneumoniae* virulence factors in host respiratory colonization and disease. Nat Rev Microbiol 2008; 6(4): 288-301.
- 82. Sempere J, Llamosí M, del Río Menéndez I, López Ruiz B, Domenech M, González-Camacho F. Pneumococcal Choline-Binding Proteins Involved in Virulence as Vaccine Candidates. Vaccines 2021; 9(2): 181.
- 83. Fathi A., Barak M, Damandan M, Amani F, Moradpour R, Khalilova I., Valizadeh M. Neonatal Screening for Glucose-6-phosphate dehydrogenase Deficiency in Ardabil Province, Iran, 2018-2019. Cell Mol Biomed Rep 2021; 1(1): 1-6.
- 84. Schachern PA, Tsuprun V, Ferrieri P, Briles DE, Goetz S, Cureoglu S, et al. Pneumococcal PspA and PspC proteins: potential vaccine candidates for experimental otitis media. Int J Pediatr Otorhinolaryngol 2014; 78(9): 1517-1521.