

Cellular and Molecular Biology

E-ISSN: 1165-158X / P-ISSN: 0145-5680



www.cellmolbiol.org

TRAIL mediated signaling in different cancers: cancer in the "Crosshairs"

Ammad Ahmad Farooqi^{1*}, Humaira Naureen², Seher Yilmaz³, Uteuliyev Yerzhan Sabitaliyevich⁴, Azamat Zhailganov⁵, Marat Rabandiyarov⁵, Ilknur Ucak⁶, Lazzat Karasholakova⁷, Rukset Attar⁸

¹Institute of Biomedical and Genetic Engineering (IBGE), Islamabad, 44000, Pakistan

² Riphah Institute of Pharmaceutical Sciences, Riphah International University, Islamabad, Pakistan

³ Department of Anatomy, Yozgat Bozok University Faculty of Medicine, Yozgat, Turkey

⁴ Department of Health Policy and Health Care Development, Kazakh Medical University of Continuing Education, Almaty 050004, Kazakhstan

⁵Department of Neurosurgery, City Children's Clinical Hospital №2, Almaty, Kazakhstan

⁶Faculty of Agricultural Sciences and Technologies, Nigde Omer Halisdemir University, Nigde, Turkey

⁷ Department of Agronomy and Technical disciplines, Zhetysu University named after Iliyas Zhansugurov, Str. I.Zhansugurov,

187A, Taldykorgan, 040009, Republic of Kazakhstan

⁸ Department of Obstetrics and Gynecology, Yeditepe University, Turkey

*Correspondence to: farooqiammadahmad@gmail.com

Received September 15, 2020; Accepted November 15, 2020; Published December 20, 2020

Doi: http://dx.doi.org/10.14715/cmb/2020.66.8.1

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Abstract: Cancer is a therapeutically challenging disease because of its heterogeneous and multifaceted nature. Decades of research have sequentially and systematically enabled us to develop a sharper and better understanding of the heterogeneous nature of cancer. Genetic, genomic and proteomic studies have unraveled wide-ranging signaling cascades which play cornerstone role in disease onset and progression. More importantly, activation of pro-survival signaling and loss of apoptosis also play critical role in cancer progression. TRAIL-mediated signaling pathway has emerged as one of the most comprehensively analyzed cascade because of its exceptional ability to target cancer cells while leaving normal cells intact. TRAIL discovery and landmark achievements related to TRAIL/TRAILreceptor signaling pathway attracted the attention of researchers. Therefore, scientists started to add missing pieces to an incomplete jig-saw puzzle and allowed contemporary researchers to conceptualize a better molecular snapshot of TRAIL-induced signaling in various cancers. Circumstantial evidence illuminated a functionally unique "push and pull" between anti-apoptotic and pro-apoptotic proteins in different cancers. Overexpression of anti-apoptotic proteins and inactivation of pro-apoptotic proteins shifted the balance towards loss of apoptosis. There has been a breakneck increase in the number of clinical trials related to TRAIL-based therapeutics which validate the true pharmacological potential of TRAIL-based therapeutics as effective anticancer agents. However, apart from advancements in our clinical understanding about the efficacy of TRAIL-based therapeutics, researchers have also faced setbacks in the field of translational medicine. Therefore, in this review, we have attempted to set spotlight on the most recent and landmark discoveries which have leveraged our understanding related to TRAIL-mediated signaling altogether to a new level.

Key words: TRAIL; Cancer; Apoptosis; Signaling.

Introduction

The removal of functionally dispensable, infected or potentially neoplastic cells is challenging and needs highly complicated and sensitive molecular mechanisms. Therefore, removal of unwanted cells is regulated by specialized and characteristically distinct programmed cell death (PCD) pathways, signifying their fundamental roles in homeostasis, host defense against pathogens, wide ranging pathologies and cancers.

Apoptosis is triggered through the activation of the apoptotic caspases and can operate either via an intrinsic or an extrinsic pathway. The intrinsic pathway is "switched on" by mitochondrial damage. Consequently, cytochrome c gets released from the mitochondrion into the cytoplasm, combines with APAF (apoptotic protease activating factor) and a pro-caspase-9 to form a signalosome known as "apoptosome" that activates caspase-9. Functionally active caspase-9 proteolytically processes and activates caspase-3, leading to cell death by cleavage of different cellular endogenous substrates.

Whereas, extrinsic pathway is activated by TRAIL, TNF, FasL which bind to death receptors (TRAIL-R, TNFR, Fas). Ligand-receptor interaction caused the oligomerization of these receptors leading to the recruitment and activation of caspase-8, which cleaved procaspase-3 to mediate apoptosis (1,2,3). Additionally, caspase-8 cleaved Bid and generated a truncated fragment, tBid. tBid translocated to the mitochondria and formed Bax/Bak pores on its surface, potentiated the release of cytochrome c and activated apoptosis (shown in figure 1). IAP (Inhibitor of apoptosis) proteins effectively blocked apoptotic death by inhibition of the activities of the caspases that executed programmed cell death. IAP protein family includes different members that have been extensively characterized. Structural studies have shown that one, two or three BIR (baculovirus IAP repeat) domains are critical for anti-apoptotic activities. Mostly IAPs have a carboxy-terminal RING domain. This domain serves as a ubiquitin ligase. Counterbalancing of IAPs is very necessary and therefore, during apoptotic death, SMAC/DIABLO (Second mitochondria-derived activator of caspase), an IAP antagonist gets released from the mitochondria that enhances the activation of caspase within the apoptosome molecular machinery. Apoptosome is formed by cytochrome c, APAF1 (apoptotic protease activating factor 1) and caspase-9. SMAC/DIABLO counteracted IAPs via four-residue IAP-binding motif present in its amino terminal region. IAPs mediated inhibition of caspase activity depends on the IAP involved. XIAP (X-linked IAP) inhibited caspase 3, caspase 7 and caspase 9 and showcased characteristically unique ability to inhibit caspases by direct binding. Series of studies have shown that BIR2 and BIR3 domains of XIAP critically regulated caspase activation. Upon activation of procaspase 9, both SMAC and XIAP compete in a mutually exclusive manner for binding to the newly exposed four residues at the amino terminal of the smaller p12 subunit of caspase 9. This mutually exclusive binding actually determined the fate of the cell in context of execution or inhibition of apoptosis.

This four-residue peptide provided pharmaceutical basis for the design of SMAC mimetics (small peptidomimetics) that duplicate the binding activity of SMAC protein to XIAP, c-IAP1 and c-IAP2. Interaction of SMAC mimetics with c-IAP1 and c-IAP2 caused conformational changes in these proteins. These changes stimulated the endogenous E3 ubiquitin ligase function of c-IAP1 and c-IAP2, which resulted in the autoubiquitination and degradation. Furthermore, SMAC mimetics prevented the binding of XIAP to caspase-3, -7, or -9. Although endogenous SMAC effectively targeted XIAP, c-IAP1 and c-IAP2 for degradation, however SMAC mimetics can further be engineered to have more specificity and efficacy.

TRAIL-mediated pathway has gained overwhelming appreciation because of its characteristically distinct ability to induce killing of the cancer cells (4,5,6). These remarkable properties captivated the attention of researchers and multiple aspects related to TRAILdriven pathway were uncovered. TRAIL-mediated apoptotic effects were explored in almost all cancers but gradually different research-groups started to note resistance against TRAIL-driven pathway. Resistance against TRAIL was puzzling and scientists started to deeply analyze the underlying mechanisms which abrogated TRAIL-driven pathway. There has been an exponential growth in the list of high-impact publication which comprehensively analyze multiple facts of TRAIL-driven pathway in different cancers. Excitingly, experts worked hard to resolve the outstanding questions related to TRAIL resistance mechanisms. Accordingly, scientists started to unlock the mystery of underlying mechanisms which caused TRAIL-resistance. It was suggested that downregulation of death receptors, overexpression of anti-apoptotic proteins and inactivation of pro-apoptotic proteins played critical role in regulation of TRAIL-mediated transduction cascade. Identification of the positive and negative regulators of TRAIL-mediated apoptotic pathway impelled researchers to analyze multiple target proteins in this network.

We have partitioned this multi-component review into sub-sections. We have summarized recently published cutting-edge researches related to TRAIL-driven pathway.

Recent breakthroughs in TRAIL-driven signaling: fresh from the pipeline

Ground-breaking discoveries in the past few years have revolutionized our understanding related to TRAIL-driven pathway.

It has recently been convincingly revealed that TRAIL-resistant sub-populations protect and enclose TRAIL-hypersensitive cells, thus causing an increase in TRAIL-resistance (7). TRAIL-resistant layers were formed at the interface of quiescent and proliferating cells and lacked both DR4 and DR5. Deprivation of nutrients and oxygen triggered an increase in the levels of DR5 in TRAIL-hypersensitive cells present within the inner spheroid layers. Cyclo-oxygenase (COX-II) inhibitors evoked DR5 expression in spheroids, most likely resulting from increased endoplasmic reticulum stress and thus re-sensitizing TRAIL-resistant cell layers to treatments (7).

Tristetraprolin, an ARE-binding protein has a critical role in the modulation of mRNA stability of death receptors (8). DR4 mRNA contained three AREs and DR5 mRNA contained four AREs in 3'-UTR. Tristetraprolin physically associated with AREs in DR4 and DR5 and enhanced the decay of DR4/5 mRNAs. Tristetraprolin overexpression in colon cancer cells enhanced TRAIL resistance but downregulation of tristetraprolin increased TRAIL sensitivity via DR4/5 expression (8).

It has previously been reported that TRAIL-resistant breast cancer cells demonstrated notably higher levels of autophagosomes (9). However, the use of autophagy inhibitors drastically reduced the number of autophagosomes. TRAIL-resistant breast cancer cells contain higher basal levels of autophagosomes. It is exciting to note that DR4 and DR5 co-localize with LC3-II within autophagosomes and upon disruption of autophagosome formation, shuttle back to the cell surface (9).

Internalization of death receptors is highly complicated. Internalized receptors can be transported from endosomes to the lysosomes where they can undergo degradation. RALB is essentially involved in trafficking of the vesicles and activation of the autophagosome assembly (10). Intriguingly, this function might be contributory to the accumulation of DR5 in RALB knockdown cancer cells. Chloroquine (lysosomal inhibitor) induced an increase in DR5 levels. Also, RALB inhibition evidently enhanced chloroquine-induced DR5 upregulation. However, chloroquine-driven increase in DR5 expression was abrogated in the cancer cells which transiently overexpressed RALB. Importantly, RALB overexpression resulted in an increased co-localization of endogenous DR5 with LAMP1. TRAIL treatment potentiated the binding of RALB with the DISC. However, RALB inhibition using pharmacologic strategies or RNAi approaches potently increased TRAIL-induced cell death in colorectal cancer cells (10).

TRAIL dose-dependently enhanced nuclear accumulation of DR4 and DR5 in pancreatic cancer cells (11). Moreover, DR4 and DR5 translocated into chromatinrich fractions. Study also provided clues about exit of death receptors. Exportin-1/chromosome region maintenance 1-homolog (CRM-1) is best-studied exporter of most nuclear proteins. CRM1 interacted with DR5 more efficiently as compared to DR4 (11).

It is interesting to note that recent publications have provided clues about nuclear accumulation of death receptors.

TRAIL-resistance mechanisms

Development of resistance against TRAIL is a major obstacle. Confluence of information pinpointed to an

ever-increasing list of proteins which played central role in making cancer cells resistant to TRAIL.

Protein O-GlcNAcylation is a hallmark post-translational modification that regulates different molecular mechanisms (12). O-GlcNAcylation enhanced TRAIL resistance, whereas O-GlcNAcylation inhibition effectively promoted TRA-8-induced apoptosis in TRAILresistant cancer cells. O-GlcNActransferase knockdown caused a notable increase in the efficacy of TRA-8 in mice xenografted with TRAIL-resistant cancer cells. Increased FADD binding to trimerized DR5 was noticed in the OGT silenced cancer cells. These findings clearly supported the concept that inhibition of O-GlcNAcylation promoted oligomerization of DR5 and FADD recruitment to form death inducing signaling complex (12).

CCR4-NOT transcription complex (CNOT), composed of 11 subunits has been shown to participate in multiple cellular processes (13). Moreover, CNOT2, one of the CCR4-NOT subunits played a central role in enhancing TRAIL-resistance. CNOT2 knockdown markedly increased TRAIL sensitivity in H1299 and A549 cells (13).

ROMO1 (Reactive oxygen species modulator-1) has also been shown to interfere with TRAIL-mediated apoptosis (14). TRAIL remarkably induced regression of tumors in mice xenografted with ROMO1-silenced HCT116 cancer cells. ROMO1 knockdown significantly lowered Bax ubiquitination by interfering with Parkin/Bax interaction (14).

EP300 gene encoded a histone acetyltransferase (p300) that regulated transcription via chromatin remodeling (15). P300 and CREBBP (CREB-binding protein) inhibition enhanced the expression levels of caspase-3, -7, -8 and 9 at the mRNA levels. Furthermore, downregulation of EP300 and CREBBP improved TRAIL-mediated apoptosis (15).

Protein phosphatase 2A (PP2A) played a critical role in TRAIL resistance (16). PP2A inhibitor LB-100 efficiently inhibited growth of the tumors in mice xenografted with MDA-468 breast cancer cells (16)

Methionine addiction is one of the hallmarks of cancer (17). Methionine restriction of methionine-addicted cancer cells increased DR5 expression. Methionine restriction increased tigatuzumab-induced activation of caspase and apoptotic death in BxPC-3 and MIA PaCa-2 cancer cells. Recombinant methioninase, a methioninedegrading enzyme is effective against different cancers. o-rMETase and tigatuzumab markedly induced regression of tumors in mice xenografted with MIA PaCa-2 cancer cells (17). Mixed lineage kinase domain-like (MLKL) also played a contributory role in the generation of extracellular and intraluminal vesicles (18). MLKL depletion heavily interfered with the intracellular trafficking of TRAIL/DR5 to degradative compartments. DR5 activation promoted rapid cleavage of substrates such as (AP2) adaptor protein 2. MLKL deficient cells revealed noteworthy increase in the cleavage of AP2 which clearly suggested that active TRAIL/DR complex promoted potent cleavage of proteins involved in clathrinmediated endocytosis (18).

Tumor cells carrying truncated O-glycans are relatively resistant to TRAIL-induced apoptosis, while cells expressing extended O-glycans are more sensitive to TRAIL (19). DR4/5 carrying normal Sialyl-T antigen formed homo-oligomers that are essential for death signaling, while truncated O-glycans attenuated the formation of homo-oligomeric complexes in Jurkat and LOX cells and promoted the hetero-oligomeric complex formation between DR5 and DcRs in LOX cells (19). Collectively, the findings suggested that extended O-glycans promoted homo-oligomerization of DR4/ DR5, which is essential for TRAIL-induced apoptosis. Truncated O-glycans prevented the homo-oligomerization of DR4/DR5 and promoted the hetero-oligomerization between DR5 with DcR2 lacking the death domain, which attenuated the death signaling.

It has previously been reported that importin β 1 transported DR5 to the nucleus (20). While, importin β 1 knockdown upregulated cell surface expression of DR5 which resulted in an increased TRAIL sensitivity. Use of importin β 1 inhibitors and agonistic anti-human DR5 (hDR5) antibody effectively reduced tumor growth in animal models (20).

JDP2 (Jun dimerization protein 2) is a bZip type transcriptional factor (21). Knockdown of JDP2 enhanced the expression of ATF4 target genes, including DR4 and DR5 in HeLa cells. Whereas, overexpression of JDP2 repressed ER stress-mediated transcriptional activation of DR5 which clearly suggested that JDP2 caused negative regulation of ATF4-mediated gene expression (21).

DR4 is O-GlcNAcylated at 424th serine residue in the death domain to transduce the death signals intracellularly (22). Data indicated that DR4 was not modified by O-GlcNAc in majority of the TRAIL-resistant cancer cell lines. Interestingly, promoting DR4 O-GlcNAcylation restored sensitivity of cancer cells to TRAIL. O-GlcNAcylation-defective DR4 neither formed DISC nor translocated to aggregated platforms for clustering of receptors (22).

IITZ-01, a lysosomotropic autophagy inhibitor enhanced TRAIL-mediated apoptotic death (23). Cbl (Casitas B-lineage lymphoma) contributed to DR5 ubiquitination and degradation. IITZ-01 protected death receptors from degradation by decreasing the stability of Cbl. IITZ-01 also potently enhanced USP9X-dependent degradation of survivin in cancer cells (23).

CUL7 ubiquitinated caspase-8 with non-K48-linked polyubiquitin chains at K215 (24). These polyubiquitin chains showcased a unique feature as they did not target the substrates for degradation by proteasomes (shown in figure 1). CUL7-mediated ubiquitination of caspase-8 and notably reduced cleavage of caspase-8 and activa-

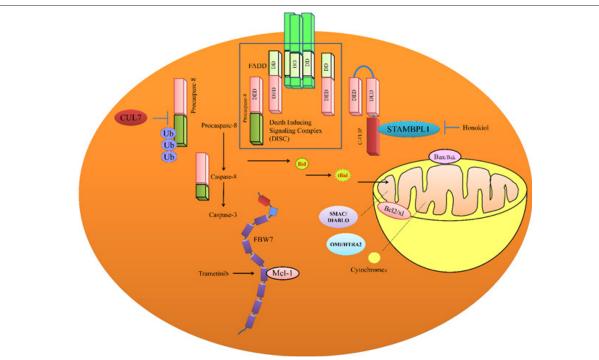


Figure 1. TRAIL-mediated intracellular signaling. Death receptor, FADD and pro-caspase-8 formed a complex for the activation of caspase-8. However, DISC formation is often disrupted by c-FLIP. Moreover, CUL7 mediated addition of non-K48-linked polyubiquitin chains to caspase-8. This ubiquitination although did not degrade caspase-8 but blocked its activation. STAMBPL1 played a dominant role in stabilization of c-FLIP. However, honokiol has been shown to interfere with STAMBPL1-mediated stabilization of c-FLIP. Trametinib induced FBW7-mediated degradation of Mcl-1.

tion. TRAIL markedly reduced tumor growth in mice inoculated with CUL7-silenced MDA-MB-231 cancer cells (24).

Glucosamine and TRAIL combinatorially suppressed the expression of c-FLIP, BCL-XL, Mcl-1 and XIAP and promoted the translocation of BAK to the mitochondrial outer membrane thereby facilitating release of the cytochrome c and SMAC (25).

HuP10 is present in the nucleus. However, excitingly, TRAIL not only induced nuclear exit of HuP10 but also promoted its entry into mitochondria (26). HuP10 is shuttled from nucleus to the cytoplasm through CRM1/ Exportin-1. Moreover, caspase-3 also played instrumental role in nuclear exit of HuP10. Findings indicated that HuP10 did not move out of the nucleus in caspase-3 knockdown cancer cells. Additionally, it was shown that HuP10 increased caspase-3 activity in cytoplasm (26).

Targeting of anti-apoptotic proteins

Bromodomain and extraterminal (BET) proteins (BRD2, BRD3, BRD4) work as epigenetic readers and master transcriptional coactivators and are now recognized cancer therapeutic targets (27). BET degraders such as ZBC260 and dBET represent a novel class of BET inhibitors that degrade BET proteins. BET degraders effectively induced apoptosis in sensitive NSCLC cells and were accompanied by reduction of Mcl-1 and c-FLIP levels. ZBC260 and TRAIL combinatorially induced apoptosis primarily through degradation of Mcl-1 and c-FLIP (27).

Trametinib, a MEK1/2 inhibitor potently degraded Mcl-1 through the proteasomal pathway (28). GSK-3β phosphorylated Mcl-1 at S159 and promoted Mcl-1 degradation. FBW7 (F-box and WD repeat domain-containing 7), a ubiquitin ligase has been shown to ubiquitinate wide-ranging substrates. FBW7 polyubiquitinated Mcl-1 and enhanced its degradation (shown in figure 1). Trametinib induced an increase in the interaction of FBW7- and Mcl-1. Mcl-1 was not ubiquitinated in FBW7 silenced cancer cells. Collectively, these findings supported the notion that trametinib induced GSK-3βmediated phosphorylation of Mcl-1 and consequent degradation by FBW7 (28).

Dexamethasone enhanced Cbl levels and interfered with TRAIL-mediated intracellular signaling. Cbl markedly induced ubiquitination of DR5, but catalytically inactive Cbl mutant did not ubiquitinate DR5 (29). Interestingly, AR-A014418 (GSK-3 β inhibitor) inhibited dexamethasone-induced Cbl stability and DR5 degradation. Furthermore, dexamethasone also induced upregulation of c-FLIP(1) by GSK-3 β (29).

Targeting of oncogenic network in TRAIL pathway

CUDC-907 is an efficient dual-acting inhibitor of PI3K (phosphoinositide 3-kinase) and HDAC (histone deacetylase) (30). CUDC-907 stimulated DR5 expression, lowered the levels of anti-apoptotic proteins Bcl-2, Bcl-xL and XIAP. DR5 knockdown abolished apoptotic death induced by the combination of CUDC-907 and TRAIL in breast cancer cells. Importantly, CUDC-907 enhanced the phosphorylation of p38 MAPK and JNK. JNK inhibition blocked CUDC-907-induced DR5 upregulation (30). Collectively, these findings suggested that CUDC-907 potentiated TRAIL-mediated apoptosis by lowering the levels of anti-apoptotic proteins and simultaneously stimulating the levels of DR5.

Natural products mediated restoration of TRAIL pathway

TRAIL-resistance has sparked an interest to search for wide-ranging bioactive chemicals from natural sources to restore apoptotic death in resistant cancers. Therefore, with the landmark achievements related to disentangling complicated web of proteins in TRAILpathway, medicinal chemists and natural product researchers contributed by identifying different natural products having premium medicinal properties and excellent ability to sensitize resistant cancer cells to TRAIL. Consequently, starting from transcriptional upregulation of death receptors, natural products have been shown to effectively stimulate extrinsic and intrinsic pathways and pharmacologically target anti-apoptotic proteins in different cancer cell lines and preclinical animal models.

Calcium/calmodulin dependent protein kinase kinase β (CaMKK β) is an upstream kinase of AMPK (AMP-activated protein kinase) (31). CaMKK β activated AMPK which consequently induced an increase in the levels of USP51 (ubiquitin specific peptidase 51). Hispidulin has been shown to trigger the activation of CaMKK β /AMPK signaling axis to enhance the stability of Bim by USP51 in Caki cells. Hispidulin and TRAIL combinatorially induced regression of tumors in mice inoculated with Caki cells (31).

Icariin stimulated the expression of DR4 and DR5 (32). Additionally, it was shown that Icariin triggered C/ EBP homologous protein (CHOP) mediated upregulation of DR5. ERK activation was found to be necessary for the upregulation of death receptors. ERK inhibition markedly suppressed Icariin-induced expression of DR4 and DR5 (32).

Pseudolaric Acid B is a diterpenoid obtained from the root barks of *Pseudolarix kaempferi* Gordon tree. It has been noted to increase TRAIL-sensitivity (33). Intraperitoneal injections of ethanolic extracts of *Pseudolarix kaempferi* markedly reduced tumor growth in mice subcutaneously injected with HN22 cells (33).

NEO214 was chemically synthesized by covalent linkage of perillyl alcohol with rolipram using a carbamate bond (34). NEO214 was tested for efficacy against intracranial athymic tumor bearing mice and intracranial immunocompetent syngeneic mice. NEO214 effectively inhibited tumor growth in mice intracranially implanted with glioblastoma cells. To analyze the pharmacological properties of NEO214 in syngeneic immunocompetent models, GL261 mouse glioma cells were intracranially implanted into C57bl/6. To confirm whether or not NEO214 stimulated CHOP and DR5 in immunocompetent brain tumor models, GL261 mouse glioma cells were implanted into the frontal cortex of syngeneic mice. Results clearly suggested that NEO214 stimulated CHOP and DR5 (34).

Honokiol enhanced TRAIL-sensitivity by lowering the levels of c-FLIP and survivin (35). Detailed studies revealed that honokiol interfered with STAMBPL1-mediated stabilization of c-FLIP and survivin. Ectopic expression of STAMBPL1 interfered with the degradation of c-FLIP and survivin by honokiol (shown in figure 1). Whereas, levels of survivin and c-FLIP were noted to be reduced in STAMBPL1-silenced in Caki cells. STAM- BPL1 directly interacted with c-FLIP and survivin and enhanced the stability of these proteins (35).

GALNT14 belongs to a large subfamily of glycosyltransferases. GALNT14-induced DR5 O-glycosylation and promoted TRAIL sensitivity of cancer cells (36). Oridonin dose- and time dependently induced an increase in the levels of GALNT-14 in Caki and A549 cells. Combinatorial treatment induced glycosylatedhigh molecular weight DR5 in A549 and Caki cells. Collectively, these findings supported the concept that DR5 glycosylation played a vital role in enhancing the TRAIL sensitivity by oridonin (36).

There is a rapid increase in the identification of natural products having remarkable ability to restore apoptosis in TRAIL-resistant cancers (37-47).

Biochemical modifications

Threonine 166 is an important phosphorylation site and studies have shown that Thr-166 phosphorylation is essential for ROS-mediated Lys-167 ubiquitination of c-FLIP protein (48). Therefore, it seems clear that generation of ROS acts as a trigger for post-translational modifications of c-FLIP. It will be paramount to analyze additional anti-apoptotic proteins tagged for degradation in oxidative stress-induced cancer cells.

HECTD3, an E3 ubiquitin ligase interacted with DED (death effector domains) of caspase-8 and ubiquitinated caspase-8 with K63-linked polyubiquitin chains (49). K63-linked polyubiquitin did not sort caspase-8 for degradation but considerably reduced activation of caspase-8 (49).

Membrane-associated RING-CH (MARCH) ubiquitin ligase family played critical role in ubiquitination of DR4. Ligase-dead MARCH-8 variant was unable to ubiquitinate DR4. Moreover, Lysine 273 was important for MARCH-8 mediated ubiquitination of DR4 (50).

KDM4A small-molecule inhibitor compound-4 (C-4) has been shown to stimulate expression of TRAIL and DR5 (51). Nuclear receptor co-repressor (NCoR)-HDAC complexes transcriptionally downregulated TRAIL and DR5. C-4 induced dissociation of KDM4A and NCoR-HDAC complexes and promoted attachment of CBP (histone acetyltransferase) to upregulate TRAIL and DR5 (51).

Xenografted mice based studies

Vemurafenib is a selective inhibitor of BRAF kinase and highly effective against anaplastic thyroid cancer cells (52). Vemurafenib potently enhanced TRAIL-induced apoptosis and induced regression of tumors in mice xenografted with 8505C cancer cells (52).

MEDI3039 is an effective multivalent DR5 agonist. It has recently been tested for efficacy against breast cancer in xenografted mice (53). MEDI3039 induced regression of the tumors in mice injected with MDA-MB-231T cancer cells. MEDI3039 was also preclinically assessed for efficacy and any possible hazardous effects at multiple doses and more injections to evaluate dose-dependent effects and to assess if more doses might prevent disease relapses. MEDI3039 not only significantly inhibited lung metastases but also inhibited growth of established experimental metastasis in xeno-

grafted mice (53).

It has been scientifically established that c-Myc played dualistic role in carcinogenesis. Elevated c-Myc activity has been reported in breast cancer cells having high metastasizing potential (54). Increased c-Myc functions are crucial for invasive outgrowths in the brain microenvironment. However, despite highly oncogenic activities, MDA-MB-231 (brain metastasizing cells) demonstrated considerably enhanced susceptibility to TRAIL-induced apoptotic death. c-Myc-depleted-MDA-MB-231 (brain metastasizing cells) exhibited reduced TRAIL sensitivity but restoration of c-MYC re-sensitized them to TRAIL-induced apoptotic death. MDA-MB-231 (brain metastasizing cells) were injected into the arterial circulation of immune-deficient mice and later mice were treated with TRAIL. TRAIL caused significant reduction in the growth of brain metastases in xenografted mice (54).

Bone seeking MDA-MB-231-cancer cells were injected into the left ventricle of anesthetized mice (55). Intra-peritoneally administered dasatinib significantly reduced number of metastases. Moreover, dasatinib promoted significantly higher relative gain and relative lower loss in the bone volume of xenografted mice. Collectively, these findings supported the fact that dasatinib prevented metastases of osteotropic MDA-MB-231 cells to the bone (55).

Co-encapsulation of the negatively charged siHSP70 and positively-charged TRAIL worked with remarkable synergy (56). Complementary activity of TRAIL and siHSP70 synergistically inhibited tumor growth in mice inoculated with A549. Furthermore, TRAIL and siHSP70 co-treatment also potently inhibited metastatic spread of cancer cells to the lungs as evidenced by noteworthy reduction in the metastatic nodules on the surface of lungs (56).

Concluding remarks

There has always been a quest to search for the anticancer agents having valuable efficacy and minimum off-target effects. Specialized killing of cancer cells is yet another stumbling block that confounds standardization of therapies. Discovery of TRAIL is regarded as a landmark in the field of molecular oncology. Branching trajectories of TRAIL-pathway have enabled researchers to not only unravel cancer killing pathways but also the strategies to therapeutically target different anti-apoptotic proteins which abrogate TRAIL-mediated apoptotic death in variety of cancers. Excitingly, anti-apoptotic proteins have been comprehensively analyzed and emerging evidence has highlighted how these proteins escaped from ubiquitination and degradation. Identification of natural and synthetic agents having minimum off-target effects and maximum efficacy has remained an overarching goal in molecular oncology. Moreover, regulation of TRAIL-induced cascade by non-coding RNAs is also a very exciting area of research and there is a gradual carving of this facet of TRAIL-pathway. More importantly, noteworthy increase in the progression of TRAIL-based therapeutics in various phases of clinical trials is indeed encouraging and promising. Therefore, keeping in view the wealth of information sequentially gathered in the past two decades will certainly help the

researchers in overcoming TRAIL-resistance pathways and inclusion of TRAIL-based therapeutics in the list of clinically approved drugs.

Restoration of apoptotic pathway is an essential aspect of molecular oncology which is evidenced by the exponentially growing number of TRAIL-pathway-related publications. More importantly, such a framework holds the potential of far-reaching breakthroughs in the treatment of cancer, for cancer biologists and of bridging the knowledge gaps in understanding how TRAILpathway operates in different cancers and their subtypes. In-depth research related to TRAIL-based therapeutics has started to gain momentum and this remarkable strength of interest will enable cross-disciplinary collaborative works and push this exciting field of research forward to get a step closer to individualized medicine.

References

1. Pitti RM, Marsters SA, Ruppert S, Donahue CJ, Moore A, Ashkenazi A. Induction of apoptosis by Apo-2 ligand, a new member of the tumor necrosis factor cytokine family. J Biol Chem. 1996 ;271(22):12687-90.

2. Walczak H, Miller RE, Ariail K, Gliniak B, Griffith TS, Kubin M, Chin W, Jones J, Woodward A, Le T, Smith C, Smolak P, Goodwin RG, Rauch CT, Schuh JC, Lynch DH. Tumoricidal activity of tumor necrosis factor-related apoptosis-inducing ligand in vivo. Nat Med. 1999;5(2):157-63.

3. Pan G, O'Rourke K, Chinnaiyan AM, Gentz R, Ebner R, Ni J, Dixit VM. The receptor for the cytotoxic ligand TRAIL. Science. 1997;276(5309):111-3.

4. Zhang XD, Franco A, Myers K, Gray C, Nguyen T, Hersey P. Relation of TNF-related apoptosis-inducing ligand (TRAIL) receptor and FLICE-inhibitory protein expression to TRAIL-induced apoptosis of melanoma. Cancer Res. 1999 ;59(11):2747-53.

5. Gliniak B, Le T. Tumor necrosis factor-related apoptosis-inducing ligand's antitumor activity in vivo is enhanced by the chemotherapeutic agent CPT-11. Cancer Res. 1999 ;59(24):6153-8.

6. Roth W, Isenmann S, Naumann U, Kügler S, Bähr M, Dichgans J, Ashkenazi A, Weller M. Locoregional Apo2L/TRAIL eradicates intracranial human malignant glioma xenografts in athymic mice in the absence of neurotoxicity. Biochem Biophys Res Commun. 1999 ;265(2):479-83.

7. Stöhr D, Schmid JO, Beigl TB, Mack A, Maichl DS, Cao K, Budai B, Fullstone G, Kontermann RE, Mürdter TE, Tait SWG, Hagenlocher C, Pollak N, Scheurich P, Rehm M. Stress-induced TRAILR2 expression overcomes TRAIL resistance in cancer cell spheroids. Cell Death Differ. 2020;27(11):3037-3052.

8. Lee WH, Han MW, Kim SH, Seong D, An JH, Chang HW, Kim SY, Kim SW, Lee JC. Tristetraprolin Posttranscriptionally Downregulates TRAIL Death Receptors. Cells. 2020 ;9(8):1851.

9. Di X, Zhang G, Zhang Y, Takeda K, Rivera Rosado LA, Zhang B. Accumulation of autophagosomes in breast cancer cells induces TRAIL resistance through downregulation of surface expression of death receptors 4 and 5. Oncotarget. 2013;4(9):1349-64.

10. Khawaja H, Campbell A, Roberts JZ, Javadi A, O'Reilly P, McArt D, Allen WL, Majkut J, Rehm M, Bardelli A, Di Nicolantonio F, Scott CJ, Kennedy R, Vitale N, Harrison T, Sansom OJ, Longley DB, Evergren E, Van Schaeybroeck S. RALB GTPase: a critical regulator of DR5 expression and TRAIL sensitivity in KRAS mutant colorectal cancer. Cell Death Dis. 2020 ;11(10):930.

11. Mert U, Adawy A, Scharff E, Teichmann P, Willms A, Haselmann V, Colmorgen C, Lemke J, von Karstedt S, Fritsch J, Trauzold A. TRAIL Induces Nuclear Translocation and Chromatin Localization of TRAIL Death Receptors. Cancers (Basel). 2019 ;11(8):1167. 12. Yang SZ, Xu F, Yuan K, Sun Y, Zhou T, Zhao X, McDonald JM, Chen Y. Regulation of pancreatic cancer TRAIL resistance by protein O-GlcNAcylation. Lab Invest. 2020;100(5):777-785.

13. Kim EO, Kang SE, Choi M, Rhee KJ, Yun M. CCR4-NOT transcription complex subunit 2 regulates TRAIL sensitivity in non-small-cell lung cancer cells via the STAT3 pathway. Int J Mol Med. 2020;45(2):324-332.

14. Jo MJ, Kim BG, Park SH, Kim HJ, Jeong S, Kim BR, Kim JL, Na YJ, Jeong YA, Yun HK, Kim DY, Han J, Heo JY, Yoo YD, Lee DH, Oh SC. Romo1 Inhibition Induces TRAIL-Mediated Apoptosis in Colorectal Cancer. Cancers (Basel). 2020 ;12(9):2358.

15. Zhang B, Chen D, Liu B, Dekker FJ, Quax WJ. A novel histone acetyltransferase inhibitor A485 improves sensitivity of nonsmall-cell lung carcinoma cells to TRAIL. Biochem Pharmacol. 2020;175:113914.

16. Uddin MH, Pimentel JM, Chatterjee M, Allen JE, Zhuang Z, Wu GS. Targeting PP2A inhibits the growth of triple-negative breast cancer cells. Cell Cycle. 2020;19(5):592-600.

17. Yamamoto J, Miyake K, Han Q, Tan Y, Inubushi S, Sugisawa N, Higuchi T, Tashiro Y, Nishino H, Homma Y, Matsuyama R, Chawla SP, Bouvet M, Singh SR, Endo I, Hoffman RM. Oral recombinant methioninase increases TRAIL receptor-2 expression to regress pancreatic cancer in combination with agonist tigatuzumab in an orthotopic mouse model. Cancer Lett. 2020 ;492:174-184.

18. Park SY, Park HH, Park SY, Hong SM, Yoon S, Morgan MJ, Kim YS. Reduction in MLKL-mediated endosomal trafficking enhances the TRAIL-DR4/5 signal to increase cancer cell death. Cell Death Dis. 2020 ;11(9):744.

19. Jiang Y, Wen T, Yan R, Kim SR, Stowell SR, Wang W, Wang Y, An G, Cummings RD, Ju T. O-glycans on death receptors in cells modulate their sensitivity to TRAIL-induced apoptosis through affecting on their stability and oligomerization. FASEB J. 2020. doi: 10.1096/fj.201900053RR.

20. Kojima Y, Nishina T, Nakano H, Okumura K, Takeda K. Inhibition of Importin β 1 Augments the Anticancer Effect of Agonistic Anti-Death Receptor 5 Antibody in TRAIL-resistant Tumor Cells. Mol Cancer Ther. 2020;19(5):1123-1133.

21. János Engler M, Mimura J, Yamazaki S, Itoh K. JDP2 is directly regulated by ATF4 and modulates TRAIL sensitivity by suppressing the ATF4-DR5 axis. FEBS Open Bio. 2020. doi: 10.1002/2211-5463.13017.

22. Lee H, Oh Y, Jeon YJ, Lee SY, Kim H, Lee HJ, Jung YK. DR4-Ser424 O-GlcNAcylation Promotes Sensitization of TRAIL-Tolerant Persisters and TRAIL-Resistant Cancer Cells to Death. Cancer Res. 2019;79(11):2839-2852.

23. Shahriyar SA, Seo SU, Min KJ, Kubatka P, Min DS, Chang JS, Kim DE, Woo SM, Kwon TK. Upregulation of DR5 and Downregulation of Survivin by IITZ-01, Lysosomotropic Autophagy Inhibitor, Potentiates TRAIL-Mediated Apoptosis in Renal Cancer Cells via Ubiquitin-Proteasome Pathway. Cancers (Basel). 2020 ;12(9):2363. 24. Kong Y, Wang Z, Huang M, Zhou Z, Li Y, Miao H, Wan X, Huang J, Mao X, Chen C. CUL7 promotes cancer cell survival through promoting Caspase-8 ubiquitination. Int J Cancer. 2019 ;145(5):1371-1381.

25. Sun C, Chesnokov V, Larson G, Itakura K. Glucosamine Enhances TRAIL-Induced Apoptosis in the Prostate Cancer Cell Line DU145. Medicines (Basel). 2019 Oct 15;6(4):104. doi: 10.3390/ medicines6040104.

26. Jana S, Hsieh AC, Gupta R. Reciprocal amplification of caspase-3 activity by nuclear export of a putative human RNA-modifying protein, PUS10 during TRAIL-induced apoptosis. Cell Death Dis. 2017 Oct 5;8(10):e3093. doi: 10.1038/cddis.2017.476. 27. Zong D, Gu J, Cavalcante GC, Yao W, Zhang G, Wang S, Owonikoko TK, He X, Sun SY. BRD4 Levels Determine the Response of Human Lung Cancer Cells to BET Degraders That Potently Induce Apoptosis through Suppression of Mcl-1. Cancer Res. 2020 ;80(11):2380-2393.

28. Lin L, Ding D, Xiao X, Li B, Cao P, Li S. Trametinib potentiates TRAIL-induced apoptosis via FBW7-dependent Mcl-1 degradation in colorectal cancer cells. J Cell Mol Med. 2020;24(12):6822-6832. 29. Jeon MY, Woo SM, Seo SU, Kim SH, Nam JO, Kim S, Park JW, Kubatka P, Min KJ, Kwon TK. Dexamethasone Inhibits TRAIL-Induced Apoptosis through c-FLIP(L) Upregulation and DR5 Downregulation by GSK3 β Activation in Cancer Cells. Cancers (Basel). 2020 ;12(10):2901.

30. Li ZJ, Hou YJ, Hao GP, Pan XX, Fei HR, Wang FZ. CUDC-907 enhances TRAIL-induced apoptosis through upregulation of DR5 in breast cancer cells. J Cell Commun Signal. 2020. doi: 10.1007/ s12079-020-00558-3.

31. Woo SM, Seo SU, Kim SH, Nam JO, Kim S, Park JW, Min KJ, Kwon TK. Hispidulin Enhances TRAIL-Mediated Apoptosis via CaMKK //AMPK/USP51 Axis-Mediated Bim Stabilization. Cancers (Basel). 2019 ;11(12):1960.

32. Kim B, Seo JH, Lee KY, Park B. Icariin sensitizes human colon cancer cells to TRAIL-induced apoptosis via ERK-mediated upregulation of death receptors. Int J Oncol. 2020;56(3):821-834.

33. Choi SJ, Ahn CH, Yang IH, Jin B, Lee WW, Kim JH, Ahn MH, Swarup N, Hong KO, Shin JA, Woo NT, Hong SD, Lee JI, Cho SD. Pseudolaric Acid B Induces Growth Inhibition and Caspase-Dependent Apoptosis on Head and Neck Cancer Cell lines through Death Receptor 5. Molecules. 2019;24(20):3715.

34. Cho HY, Thein TZ, Wang W, Swenson SD, Fayngor RA, Ou M, Marín-Ramos NI, Schönthal AH, Hofman FM, Chen TC. The Rolipram-Perillyl Alcohol Conjugate (NEO214) Is A Mediator of Cell Death through the Death Receptor Pathway. Mol Cancer Ther. 2019;18(3):517-530.

35. Woo SM, Seo SU, Kubatka P, Min KJ, Kwon TK. Honokiol Enhances TRAIL-Mediated Apoptosis through STAMBPL1-Induced Survivin and c-FLIP Degradation. Biomolecules. 2019;9(12):838.

36. Jeon MY, Seo SU, Woo SM, Min KJ, Byun HS, Hur GM, Kang SC, Kwon TK. Oridonin enhances TRAIL-induced apoptosis through GALNT14-mediated DR5 glycosylation. Biochimie. 2019;165:108-114.

37. Surapally S, Jayaprakasam M, Verma RS. Curcumin augments therapeutic efficacy of TRAIL-based immunotoxins in leukemia. Pharmacol Rep. 2020;72(4):1032-1046.

38. Nazim UM, Yin H, Park SY. Downregulation of c-FLIP and upregulation of DR-5 by cantharidin sensitizes TRAIL-mediated apoptosis in prostate cancer cells via autophagy flux. Int J Mol Med. 2020;46(1):280-288.

39. Kwon OS, Jung JH, Shin EA, Park JE, Park WY, Kim SH. Epigallocatechin-3-Gallate Induces Apoptosis as a TRAIL Sensitizer via Activation of Caspase 8 and Death Receptor 5 in Human Colon Cancer Cells. Biomedicines. 2020 ;8(4):84.

40. Woo SM, Min KJ, Kwon TK. Magnolol Enhances the Therapeutic Effects of TRAIL through DR5 Upregulation and Downregulation of c-FLIP and Mcl-1 Proteins in Cancer Cells. Molecules. 2020 ;25(19):4591.

41. Elmallah MIY, Cogo S, Constantinescu AA, Elifio-Esposito S, Abdelfattah MS, Micheau O. Marine Actinomycetes-Derived Secondary Metabolites Overcome TRAIL-Resistance via the Intrinsic Pathway through Downregulation of Survivin and XIAP. Cells. 2020;9(8):1760.

42. Zhou X, Zijlstra SN, Soto-Gamez A, Setroikromo R, Quax WJ. Artemisinin Derivatives Stimulate DR5-Specific TRAIL-Induced Apoptosis by Regulating Wildtype P53. Cancers (Basel). 2020 ;12(9):2514. 43. Lohberger B, Bernhart E, Stuendl N, Glaenzer D, Leithner A, Rinner B, Bauer R, Kretschmer N. Periplocin mediates TRAILinduced apoptosis and cell cycle arrest in human myxofibrosarcoma cells via the ERK/p38/JNK pathway. Phytomedicine. 2020 ;76:153262.

44. Peng X, He Y, Gao Y, Duan F, Chen J, Ruan H. Cytochalasins from an endophytic fungus Phoma multirostrata XJ-2-1 with cell cycle arrest and TRAIL-resistance-overcoming activities. Bioorg Chem. 2020;104:104317.

45. Cho HD, Gu IA, Won YS, Moon KD, Park KH, Seo KI. Auriculasin sensitizes primary prostate cancer cells to TRAIL-mediated apoptosis through up-regulation of the DR5-dependent pathway. Food Chem Toxicol. 2019;126:223-232.

46. Sophonnithiprasert T, Mahabusarakam W, Watanapokasin R. Artonin E sensitizes TRAIL-induced apoptosis by DR5 upregulation and cFLIP downregulation in TRAIL-refractory colorectal cancer LoVo cells. J Gastrointest Oncol. 2019;10(2):209-217.

47. Dai H, Jiang Y, Luo Y, Bie P, Chen Z. Triptolide enhances TRAIL sensitivity of pancreatic cancer cells by activating autophagy via downregulation of PUM1. Phytomedicine. 2019;62:152953.

48. Wilkie-Grantham RP, Matsuzawa S, Reed JC. Novel phosphorylation and ubiquitination sites regulate reactive oxygen speciesdependent degradation of anti-apoptotic c-FLIP protein. J Biol Chem. 2013 ;288(18):12777-90.

49. Li Y, Kong Y, Zhou Z, Chen H, Wang Z, Hsieh YC, Zhao D, Zhi X, Huang J, Zhang J, Li H, Chen C. The HECTD3 E3 ubiquitin ligase facilitates cancer cell survival by promoting K63-linked poly-ubiquitination of caspase-8. Cell Death Dis. 2013 ;4(11):e935.

50. van de Kooij B, Verbrugge I, de Vries E, Gijsen M, Montserrat V, Maas C, Neefjes J, Borst J. Ubiquitination by the membrane-associated RING-CH-8 (MARCH-8) ligase controls steady-state cell

surface expression of tumor necrosis factor-related apoptosis inducing ligand (TRAIL) receptor 1. J Biol Chem. 2013 ;288(9):6617-28.

51. Wang J, Wang H, Wang LY, Cai D, Duan Z, Zhang Y, Chen P, Zou JX, Xu J, Chen X, Kung HJ, Chen HW. Silencing the epigenetic silencer KDM4A for TRAIL and DR5 simultaneous induction and antitumor therapy. Cell Death Differ. 2016;23(11):1886-1896.

52. Pilli T, Cantara S, Marzocchi C, Pacini F, Prabhakar BS, Castagna MG. Vemurafenib may overcome TNF-related apoptosis-inducing ligand (TRAIL) resistance in anaplastic thyroid cancer cells. Endocrine. 2020;67(1):117-123.

53. Greer YE, Gilbert SF, Gril B, Narwal R, Peacock Brooks DL, Tice DA, Steeg PS, Lipkowitz S. MEDI3039, a novel highly potent tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) receptor 2 agonist, causes regression of orthotopic tumors and inhibits outgrowth of metastatic triple-negative breast cancer. Breast Cancer Res. 2019;21(1):27.

54. Lee HY, Cha J, Kim SK, Park JH, Song KH, Kim P, Kim MY. c-MYC Drives Breast Cancer Metastasis to the Brain, but Promotes Synthetic Lethality with TRAIL. Mol Cancer Res. 2019;17(2):544-554.

55. Heilmann T, Rumpf AL, Roscher M, Tietgen M, Will O, Gerle M, Damm T, Borzikowsky C, Maass N, Glüer CC, Tiwari S, Trauzold A, Schem C. Dasatinib prevents skeletal metastasis of osteotropic MDA-MB-231 cells in a xenograft mouse model. Arch Gynecol Obstet. 2020;301(6):1493-1502.

56. Zhou A, Du J, Jiao M, Xie D, Wang Q, Xue L, Ju C, Hua Z, Zhang C. Co-delivery of TRAIL and siHSP70 using hierarchically modular assembly formulations achieves enhanced TRAIL-resistant cancer therapy. J Control Release. 2019; 304:111-124.