



Bioactive compounds and health benefits of edible *Rumex* species-A review

Abhay Prakash Mishra^{1,2}, Mehdi Sharifi-Rad³, Mohammad Ali Shariati⁴, Yahia N. Mabkhot^{5*}, Salim S. Al-Showiman⁵, Abdur Rauf⁶, Bahare Salehi^{7,8*}, Milan Župunski⁹, Majid Sharifi-Rad¹⁰, Poonam Gusain¹¹, Javad Sharifi-Rad^{12,13*}, Hafiz Ansar Rasul Suleria¹⁴, Marcello Iriti^{15*}

¹ Department of Pharmaceutical Chemistry, H.N.B. Garhwal (A Central) University Srinagar Garhwal, 246174, Uttarakhand, India

² Kursk State Agricultural Academy, Kursk, 305021, Russia

³ Department of Medical Parasitology, Zabol University of Medical Sciences, Zabol, Iran

⁴ Head of Research Department, LLC «Science & Education», Russia

⁵ Department of Chemistry, College of Science, King Saud University, P.O. Box 2455, Riyadh 1451; Kingdom of Saudi Arabia

⁶ Department of Chemistry, University of Swabi, Anbar-23561, Khyber Pakhtunkhwa, Pakistan

⁷ Medical Ethics and Law Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁸ Student Research Committee, Shahid Beheshti University of Medical Sciences, 22439789 Tehran, Iran

⁹ University of Novi Sad, Faculty of Sciences, Department of Biology and Ecology, Trg Dositeja Obradovića 3, Novi Sad, Serbia

¹⁰ Department of Range and Watershed Management, Faculty of Natural Resources, University of Zabol, Zabol, Iran

¹¹ Regional Science Center, Uttarakhand State Council for Science and Technology, Dehradun 248007, India

¹² Phytochemistry Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

¹³ Department of Chemistry, Richardson College for the Environmental Science Complex, The University of Winnipeg, Winnipeg, MB, Canada

¹⁴ UQ School of Medicine, University of Queensland, Australia

¹⁵ Department of Agricultural and Environmental Sciences, Milan State University, Milan, Italy

Correspondence to: yahia@ksu.edu.sa; bahar.salehi007@gmail.com; javad.sharifrad@gmail.com

Received November 21st, 2017; **Accepted** February 2nd, 2018 ; **Published** June 25, 2018

Doi: <http://dx.doi.org/10.14715/cmb/2018.64.8.5>

Copyright: © 2018 by the C.M.B. Association. All rights reserved.

Abstract: Medicinal and food plants as well as their bioactive fractions have been used by diverse human cultures since ancient times. These plants provide multiple health benefits because of the presence of a plethora of phytochemicals including phenylpropanoids, isoprenoids, alkaloids, sulphated compounds, peptides and polysaccharides that are responsible for various biological activities such as anticancer, antioxidant, antifungal, antibacterial, anti-dysenteric, anti-inflammatory, antiulcer, anti-hypertensive and anticoagulant properties. The genus *Rumex* includes edible and medicinal herbs belonging to buckwheat (*Polygonaceae*) family, consisting of about 200 species rich in phenylpropanoids and anthraquinones. Some *Rumex* species have exhibited health-promoting effects and have been used as traditional foods and herbal remedies, though a limited information has been documented on their specific biological properties. Therefore, this survey aimed at reviewing the *Rumex* species with documented biological activity, focusing on preclinical evidences on their efficacy and safety.

Key words: *Polygonaceae*; Buckwheat; Functional foods; Nutraceuticals; Traditional foods; Herbal remedies.

Introduction

Medicinal plants can be a promising alternative for many diseases and conditions (1-17). Always, these plants are also valued to flavor foods, giving the food a dual role, i.e. flavor and bioactive compounds (6, 18-23). Furthermore, medicinal plants are low cost and tend to have fewer side effects than synthetic drugs (21, 24-27).

The family *Polygonaceae* comprises about 50 genera and 1200 species that are worldwide distributed. Important genera include *Rheum*, *Rumex*, *Polygonum*, *Coccoloba*, *Calligonum* and *Persicaria* (28), traditionally used as herbal remedies for treating several ailments including urinary inflammation, gallstones, chronic cutaneous diseases, skin burns, hepatitis, jaundice, fever, osteomyelitis and as anticancer, diuretic and laxative agents (29-37). Knotweed (*Fallopia japonica*) has been used traditionally in Asia for treating hepatitis, inflammation, skin burns, osteomyelitis and gallstones (38).

Roots of *Rumex crispus* have been applied in folk medicine to cure jaundice, fever, constipation and chronic cutaneous diseases, whereas fruits and seeds have been prescribed for treating hepatitis and cancer. Rhizome of *Polygonum sachalinense* has been used in oriental medicine as diuretic and laxative (39, 40).

As other plant families, *Polygonaceae* are rich in secondary metabolites, in particular phenylpropanoids and anthraquinones, possibly responsible for the health properties ascribed to these plant species (29).

Genus *Rumex*

About 250 species are included in the genus *Rumex*, both annual and perennial herbs worldwide distributed. Previous studies have reported anticancer, antidiarrheal, antioxidant, analgesic, anti-inflammatory, anthelmintic and antimicrobial activities of plants belonging to this genus (40), rich in bioactive phytochemicals (Figure 1).

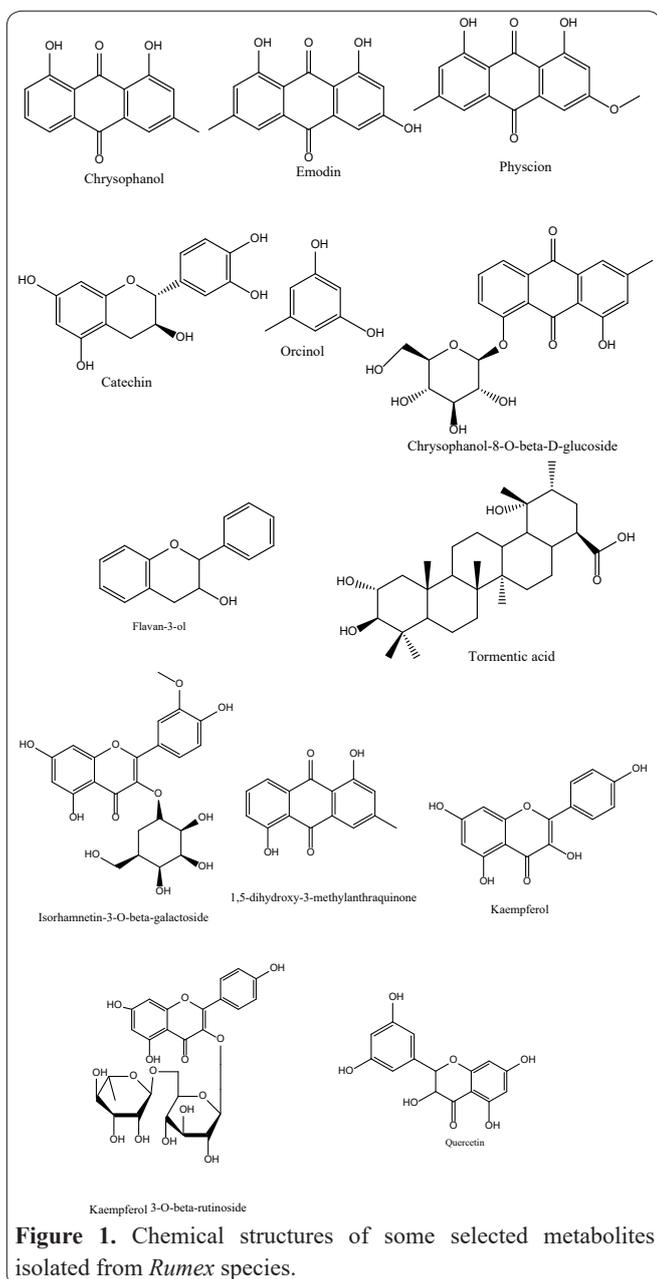


Figure 1. Chemical structures of some selected metabolites isolated from *Rumex* species.

Chemical and pharmacological profiles of genus *Rumex*

Kang *et al.* (41) investigated crude methanol extract obtained from roots of *R. gmelinii* and isolated 1-*O*- β -D-glucopyranosyl chrysophanol and 1-*O*- β -D-glucopyranosyl emodin. Demirezer *et al.* (42) reported 9 compounds from roots of *R. patientia* including chrysophanol, physcion, catechin, emodin, flavan-3-ol, 6-chlorocatechin, chrysophanol-8-*O*- β -D-glucopyranoside, emodin-8-*O*- β -D-glucopyranoside and orcinol. Among the isolated compounds, flavan-3-ol, 6-chlorocatechin and catechin exhibited antiradical scavenging activity. Kim *et al.* (43) reported ethyl gallate, 4(R),23-epoxy-2,3,19-trihydroxy-24-norurs-12-en-28-oic acid, 2,3,19-trihydroxy-24-norurs-4(23),12-dien-28-oic acid, tormentic acid and myrianthic acid from ethyl acetate soluble extract of the stem of *R. japonicus*. The 2,3,19-trihydroxy-24-norurs-4(23),12-dien-28-oic acid and ethyl gallate showed a significant inhibitory activity on AGEs (advanced glycation end products) formation with IC₅₀ values of 87 and 14.3 μ g/ml on RLAR (rat lens aldose reductase), respectively. Bařkan *et al.* (44)

isolated 1,5-dihydroxy-3-methylanthraquinone, 1,3,5-trihydroxymethylanthraquinone and 1,5-dihydroxy-3-methoxy-7-methylanthraquinone from roots of *R. crispus*.

Kim *et al.* (45) reported ω -hydroxyemodin, emodin, chrysophanol-8-*O*- β -D-glucoside, physcion-8-*O*- β -D-glucoside and emodin-8-*O*- β -D-glucoside. They also isolated five flavonoids: quercetin, kaempferol-3-*O*- β -D-glucoside, isoquercitrin and (+)-catechin from fruits of *R. japonicus* and evaluated their aldose reductase inhibitory potential. Wang *et al.* (46) isolated two oxanthrones C-glucoside, 6-methoxyl-10-hydroxyaloin A and 6-methoxyl-10-hydroxyaloin B from roots of *R. gmelinii*.

Hawas *et al.* (47) have isolated kaempferol 3-*O*- β -galactoside, kaempferol 3-*O*- β -glucoside, kaempferol 3-*O*-rutinoside, isorhamnetin 3-*O*- β -galactoside, isorhamnetin 3-*O*- β -glucoside, isorhamnetin 3-*O*-rutinoside from methanolic extract of *R. dentatus*. These compounds exhibited moderate antimicrobial activity, weak antioxidant and cytotoxic activities. Jo *et al.* (48) reported 1,8-dihydroxy-3-methoxy-6-methylanthracene-9,10-dione isolated from roots of *R. japonicus* and its antitumor activity. Ahmed *et al.* (49) investigated urease inhibitory potential of crude methanol extract of *R. acetosella* roots and its sub-fractions including *n*-hexane, chloroform, ethyl acetate, *n*-butanol and aqueous fraction.

Functional foods

Food habits and trends in food production and consumption have health, environmental and social impacts. Functional foods are being considered as a ‘magic food’ to alleviate some of the health care costs associated with aging. One of such example is naturally occurring fatty acids found in milk fat and adipose tissue of ruminants, whose quantity in food products can be enhanced by feeding animals a specialized diet (50, 51). “A food can be regarded as ‘functional’ if it is satisfactorily demonstrated to affect beneficially one or more target functions in the body, beyond adequate nutritional effects, in a way that is relevant to either an improved state of health and well-being and/or reduction of risk of disease” (52).

Phenolic compounds are plant secondary metabolites, which play an important role in defense against pathogens and pests, as well as in species dissemination. The interest in these compounds is related with their capacity to counteract oxidative stress involved in the pathogenesis of more than 100 diseases including malaria, atherosclerosis, cancer, diabetes, acquired immunodeficiency syndrome, thus promoting health benefits. Various bioactive phytochemicals from plants used as functional foods in specific disorders are summarized in Table 1. Jimoh *et al.* (53) studied polyphenolic content and antioxidant and antibacterial activities of the acetone, methanol, and water extracts of *Rumex ecklonianus* plant. A comparative study between wild edible plants including *Rumex* spp. and *Cirsium pumilum* showed that *Rumex* spp. contained higher amount of total phenols (102.56 \pm 3.13mg/100g) compared with *Cirsium pumilum* (93.64 \pm 0.28mg/100g)(54).

As previously introduced, *Rumex* is one of the most

Table 1. Functional foods and their health benefits.

Component	Product	Source	Health benefits	References
Flavones	Flax seed	<i>Linum usitatissimum</i>	Neutralize free-radicals and reduce risk of cancer.	
β -Glucan, vitamins, minerals and antioxidants	Oats	<i>Hordeum vulgare</i>	Reduce risk of cardiovascular disease, lower LDL and total cholesterol.	
Omega-3 fatty acids	Fish oils	Salmon	Reduce risk of cardiovascular disease and improve mental functions.	
Catechins	Tea	<i>Camellia sinensis</i>	Neutralize free radicals and reduce risk of cancer.	
Lycopene	Tomato products	<i>Solanum lycopersicum</i>	Reduces the risk of prostate cancer.	(65)
Lactic acid	Yogurt	<i>Lactobacillus</i> spp.	Improves quality of intestinal microflora.	
Pro-vitamin A	Golden rice, orange-fleshed sweet potato	<i>Oryza sativa</i> , <i>Ipomoea batatas</i>	Supports the immune system, strengthens vision, assists with normal organ function and reproduction.	
Xylitol	Chewing gum	Berries, oats, and mushrooms	Prevents dental caries.	
Sterols	Margarine	Plants	Improve cholesterol levels.	

important genera belonging to Polygonaceae distributed worldwide (European, Asian, African and American countries). Approximately 200 species of this genus have been reported to possess culinary as well as medicinal uses for the treatment of pain, inflammation, bleeding, tinea, tumour and constipation in Ayurveda, Siddha, Unani and Chinese healing systems (Table 2). The roots of *Rumex hastatus* are of medicinal significance for cough, headache and fever. *Rumex* species are rich in anthraquinones, naphthalenes, flavonoids, stilbenoids, triterpenes, carotenoids and phenolic acids (Figure 1). Previous studies revealed the presence of sterols in *R. nepalensis* and anthraquinones in *R. hastatus* (30, 55).

Economic value and relevance for functional food area

Traditional use of different *Rumex* species has taken places in different parts of the world. It has historical background along with the potential to continue contributing much in the future by supporting sustainable development of societies and economies. Interrelationship among the past, the present and the future is woven in human cultures and their habits to use plants for different purposes such as medicine, food, pharmaceuticals etc. (31). According to the traditional knowledge, these species have been used in medical practice, both in ethnoveterinary and ethnomedicine for improving of health and welfare (32-34, 56, 57). Usefulness for human diet is documented in numerous studies. For example, eleven *Rumex* species are widely used across Balkan region for preparation of traditional dishes (58). In Belarus, amongst different wild edible species there are currently in use three *Rumex* spp. for cooking, particularly in soups (59). Traditionally, in Palestinian region the leaves of *Rumex acetosa* are used as filling for a traditional pie called sambossek, or fried in olive oil and eaten (60). Up to four *Rumex* species are sold at the markets in Morocco as green leafy vegetables (61). In Mediterranean parts of Croatia, these species are usually sold at the markets as a part of wild leafy vegetable

mixes. As authors have stated, the use of these mixes is much less widespread in northern parts of the country, possible due to the ecological-economical explanation - with lack of the arable land they have to eat wild products. The price of such leafy vegetable mixture is 1.6-3.2 USD per kg as reported by Łuczaj *et al.* (62), with sold up to 4 kilograms of vegetable mixture at least once per week. Such data are promising and point to the beneficial aspect of collecting and cultivating these species. In the study conducted by Kasper-Pakosz *et al.* (63), it is reported that amongst different plant species being sold at the markets in Poland, *Rumex acetosa* is present as well. Further, authors have stated that most of the sold plants are also cultivated and not only harvested from native habitats. Such practice is promising from the economical point of view, conservation strategy and implementation of functional food in daily diet and eating habits. As reported by Łuczaj *et al.* (64) many young people have oriented to the new trends in nutrition and self-medication in Europe, mostly thanks to the spreading of information via the Internet. Intensive production (cultivation) of these and other wild species is not possible if food habits will not change. In this view, economic importance and benefits of markets are influencing the process of cultural transmission of traditional uses of food plants. The main issue is scarcity of the available data from open markets in different countries that could give an insight into the offered wild herbs. Due to their medicinal properties and healthy potential, they are suitable candidate for functional food area. Rich in bioactive components, freshly eaten or cooked, wild herbs are promising vegetables for vegetarian diets and omnivores as well. Their presence on market stands in different countries, perseverance through traditional knowledge and richness in bioactive phytochemicals indicate the rising needs for functional food area, even if collecting of wild herbs is considered by some people time-consuming and season-dependent (64). Economic importance might be well correlated with increasing interest for functional food area worldwide, which further may lead to intensified production of these species and their presence at market stands.

Table 2. Bioactive phytochemicals, traditional uses, parts used and biological activities of *Rumex* spp. plants.

Plant species	Bioactive compounds	Medicinal/commercial uses	Parts used	Biological activities	References
<i>Rumex alpinus</i>	2-Acetyl-3-methylnaphthalene-1,8-diol.	Laxative, jaundice, astringent, constipation, diarrhoea, eczema.	Leaves, roots.	Antibacterial.	(66-68)
<i>Rumex obtusifolius</i>	Anthracene derivatives, flavonoids, procyanidins, oxalic acid.	Burns and boils, blisters, nettle stings/wrapping up butter, sores, tumors, hepatic, eye and dermatitis, furuncles, bruises, jaundice and fever.	Leaves, roots.	Antidote, depurative, astringent, laxative, antioxidant and antibacterial, disinfectant, scar healer and as anti-arthritic and anti-anemic tonic.	(68-70)
<i>Rumex aquaticus</i>	Anthraquinones (emodin, chrysophanol, physcion, citreorosein, chrysophanol-8-O-glucoside), flavonoids (quercetin, quercetin-3,3'-dimethylether, isokaempferide, quercetin 3-O-arabioside, quercetin 3-O-galactoside, quercetin 3-O-glucoside catechin), stilbenes (resveratrol, piceid) and 1-stearoylglycerol.	Astringent, tonic, diarrhoea, ulcers, edema, disinfestation, jaundice, antipyretic, neuroprotective. Alleviation of inflammation in the gastrointestinal tract, preventing H ₂ O ₂ -induced cytotoxicity through increasing cell viability and reducing ROS production.	Leaves, roots, seeds.	Neuroprotective. Cytoprotective effects against H ₂ O ₂ -induced oxidative stress, enhancement of the cellular antioxidant capacity.	(35, 68, 71, 72)
<i>Rumex crispus</i>	Rumicin, chrysarobin, β -sitosterol, hexadecanoic acid, hexadecanoic-2,3-dihydroxy propyleste, chrysophanol, physcion, emodin, chrysophanol-8-O- β -D-glucopyranoside, physcion-8-O- β -D-glucopyranoside, emodin-8-O- β -D-glucopyranoside, gallic acid, (+)-catechin, kaempferol, quercetin, kaempferol-3-O- α -L-rhamnopyranoside, quercetin-3-O- α -L-rhamnopyranoside.	Hemorrhage and dermolysis. Laxative, tonic action, rheumatism, bilious complaints, astringent in piles, bleedings of the lungs, spring eruption, scurvy, scrofula, jaundice, bowels, cancer, diphtheria. Food as soups, sauces and salads. Young leaves are used to prepare "dolma" with minced meat or roasted to prepare a meal. Suppression of RANKL-induced trabecular bone loss by preventing microstructural deterioration.	Leaves, roots, seeds.	Protection against osteoporosis, possible increase in osteoblast differentiation. Trabecular bone loss by preventing microstructural deterioration.	(36, 68, 73, 74)
<i>Rumex hastatus</i>	Leucodelphinidin, leucopelargonidin.	Anticancer.	Stems, leaves, roots.	Antioxidant, antimycobacterial in skin infections, gastrointestinal disturbances.	(75, 76)
<i>Rumex patientia</i>	Emodin-6-O- β -D-glucopyranoside, flavan-3-ol, 6-chlorocatechin, 2-acetyl-3-methyl-6-carboxy-1,8-dihydroxynaphthalene-8-O- β -D-glucopyranoside, labadoside (4,4''-binaphthalene-8,8''-O,O-di- β -D-glucopyranoside), orientaloid (2-acetyl-3-methyl-1,8-dihydroxynaphthalene-8-O- β -D-glucopyranoside), patientosides A and B.		Leaves	Antipyretic, anti-inflammatory, analgesic, cytotoxic.	(42, 76, 77)
<i>Rumex vesicarius</i>	Flavonoids, C-glycosides: vitexin, isovitexin, orientin, iso-orientin; anthraquinones: emodin, chrysophanol, rumicin, lapathin; oxalic acid, tannins, mucilage, mineral salts and vitamin C.	Tonic, Diuretic, antiscorbutic, appetizer, laxative, astringent, carminative, stomachic and for jaundice.	Stems, leaves, roots	Antioxidant, Antimycobacterial in skin infections, gastrointestinal disturbances.	(76, 77)

<i>Rumex dentatus</i>	Helonioside A, gallic acid, isovanillic acid, p-hydroxycinnamic acid, succinic acid, n-butyl-β-D-fructopyranoside, quercetin, hexadecanoic acid 2,3-dihydroxy propyl ester, β-sitosterol, daucosterol, anthraquinones, flavonoids, phytosterols, phytosteryl esters, free fatty acids, chromones, anthrones, kaempferol 3-O-β-galactoside, kaempferol 3-O-β-glucoside, kaempferol 3-O-rutinoside, isorhamnetin 3-O-β-galactoside, isorhamnetin 3-O-β-glucoside, isorhamnetin 3-O-rutinoside, chlorogenic acid, myricetin, vitamin C.	Inhibition of proliferation of breast cancer cells; tooth extraction	Whole plant, leaves.	Antimicrobial, cytotoxic, antioxidant, breast cancer prevention and/or treatment.	(47, 76, 78, 79)
<i>Rumex japonicus</i>	Emodin, rutin, rumejaposide, epoxynaphthoquinol, chrysophanol, physcion, 8-O-β-glucopyranoside.	Hemorrhage, wounds. Decrease of the releases of pro-inflammatory cytokines and down-regulating the TLR4 and TLR2 expressions. Infections, malignant sore, constipation, tumors.	Roots, whole plant.	Antioxidant, anticancer, anti-proliferative, antimicrobial, apoptosis. Protective effect against sepsis in mice caused by lethal dose of heat killed <i>E. coli</i> .	(37, 76, 77, 80, 81)
<i>Rumex scutatus</i>		Fried in butter and then prepared as omelette.			(74)
<i>Rumex abyssinicus</i>	Rhein, chrysophanol, emodin, emodic acid, aloemodin, alizarin, physcion, damnacanthal, catenarin, anthraquinone, plamidin C, chrysophanol-8-β-D-glucoside, emodin-8-β-D-glucoside.	Depurative, haemostatic, gonorrhoea, pulmonary tuberculosis and leprosy. Cooked alone or together with other vegetables.	Shoots, leaves	Antiviral, anticancer, antibacterial, anti-inflammatory.	(82, 83)
<i>Rumex vesicarius</i>	8-C-Glucosyl-apigenin, 8-C-glucosyl-luteolin, 6-C-hexosyl-quercetin, 3-O-rutinosyl-quercetin, 7-O-rhamno-hexosyl-diosmetin, 7-O-rhamno-acetylhexosyl-diosmetin, catechin, epicatechin, ferulohexoside, 6-C-glucosyl-naringenin, epicatechin gallate, 6-C-glucosyl-catechin, epigallocatechin gallate.	Stomach heat, toothache, nausea, appetizer, jaundice, constipation, indigestion, dysentery.	Seeds	Diuretic, hepatoprotective, sedative, aperient.	(84, 85)
<i>Rumex nepalensis</i>	Rumexneposide A, physcion, chrysophanol-8-O-β-D-glucopyranoside, torachryson, emodin-8-O-β-D-glucopyranoside, emodin-8-O-β-D-(6-O-acetyl) glucopyranoside, chrysophanol, emodin, citreorosein, resveratrol, nepodin-8-O-β-D-glucopyranoside, torachryson-8-O-β-D-glucopyranoside, chrysophanol-8-O-β-D-(6-O-acetyl) glucopyranoside	Pain, inflammation, bleeding, tinea, tumor, constipation.			(55)

Conclusions

Medicinal plants have been used for thousands of years as food, feed and medicaments to treat and prevent various infectious and non-infectious diseases (86). Plants belonging to *Polygonaceae* family including *Rumex* species have been used in traditional medicine for treating several disorders including urinary inflammation, hepatitis, chronic cutaneous diseases, jaundice, fever, skin burns, osteomyelitis, gallstones and as diuretic, laxative and anticancer agents. Therefore, in this survey, we collected the latest literature on bioactives of *Rumex* species that exhibited pharmacological activities and can be suggested as innovative food and feed.

Acknowledgments

This project was supported by King Saud University, Deanship of Scientific Research, College of Sciences Research Center, and the Vice-chancellor for Research Affairs of Shahid Beheshti University of Medical Sciences, Tehran, Iran.

Conflict of interest

The authors declare no conflict of interest.

References

- Sharifi-Rad M, Varoni EM, Salehi B et al. Plants of the Genus *Zingiber* as a Source of Bioactive Phytochemicals: From Tradition to Pharmacy. *Molecules* 2017; 22(12): 2145.
- Sharifi-Rad J, Salehi B, Stojanović-Radić ZZ et al. Medicinal plants used in the treatment of tuberculosis-Ethnobotanical and ethnopharmacological approaches. *Biotechnology Advances* 2017; doi: 10.1016/j.biotechadv.2017.07.001.
- Sahraie-Rad M, Izadyari A, Rakizadeh S, Sharifi-Rad J. Preparation of strong antidandruff shampoo using medicinal plant extracts: a clinical trial and chronic dandruff treatment. *Jundishapur Journal of Natural Pharmaceutical Products* 2015; 10(4): e21517.
- Sharifi-Rad J, Sureda A, Tenore GC et al. Biological activities of essential oils: From plant chemoeology to traditional healing systems. *Molecules* 2017; 22(1): 70.
- Sharifi-Rad J, Mnayer D, Tabanelli G et al. Plants of the genus *Allium* as antibacterial agents: From tradition to pharmacy. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 57-68.
- Sharifi-Rad M, Tayeboon G, Miri A et al. Mutagenic, antimutagenic, antioxidant, anti-lipoxygenase and antimicrobial activities of *Scandix pecten-veneris* L. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(6): 8-16.
- Salehi B, Zucca P, Sharifi-Rad M et al. Phytotherapeutics in cancer invasion and metastasis. *Phytotherapy Research* 2018; doi:10.1002/ptr.6087.
- Snow Setzer M, Sharifi-Rad J, Setzer WN. The search for herbal antibiotics: An in-silico investigation of antibacterial phytochemicals. *Antibiotics* 2016; 5(3): 30.
- Sharifi-Rad M, Iriti M, Gibbons S, Sharifi-Rad J. Anti-methicillin-resistant *Staphylococcus aureus* (MRSA) activity of Rubiaceae, Fabaceae and Poaceae plants: A search for new sources of useful alternative antibacterials against MRSA infections. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 39-45.
- Sharifi-Rad J, Soufi L, Ayatollahi S et al. Anti-bacterial effect of essential oil from *Xanthium strumarium* against shiga toxin-producing *Escherichia coli*. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 69-74.

- Sharifi-Rad J. Herbal Antibiotics: Moving back into the mainstream as an alternative for "Superbugs". *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 1-2.
- Sharifi-Rad M, Tayeboon G, Sharifi-Rad J, Iriti M, Varoni E, Razazi S. Inhibitory activity on type 2 diabetes and hypertension key-enzymes, and antioxidant capacity of *Veronica persica* phenolic-rich extracts. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(6): 80-85.
- Choudhary MI, Hussain A, Adhikari A et al. Anticancer and α -chymotrypsin inhibiting diterpenes and triterpenes from *Salvia lerifolia*. *Phytochemistry Letters* 2013; 6(1): 139-143.
- Moradkhani S, Ayatollahi AM, Ghanadian M, Moin MR, Razavizadeh M, Shahlaei M. Phytochemical Analysis and Metal-chelation Activity of *Achillea tenuifolia* Lam. *Iranian Journal of Pharmaceutical Research* 2012; 11(1): 177.
- Stojanović-Radić Z, Pejčić M, Stojanović N, Sharifi-Rad J, Stanković N. Potential of *Ocimum basilicum* L. and *Salvia officinalis* L. essential oils against biofilms of *P. aeruginosa* clinical isolates. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 27-32.
- Salehi B, Mishra AP, Shukla I et al. Thymol, thyme and other plant sources: health and potential uses. *Phytotherapy Research* 2018 ; doi: 10.1002/ptr.6109.
- Sharifi-Rad M, Mnayer D, Flaviana Bezerra Morais-Braga M et al. *Echinacea* plants as antioxidant and antibacterial agents: From traditional medicine to biotechnological applications. *Phytotherapy Research* 2018; doi: 10.1002/ptr.6101.
- Sharifi-Rad J, Salehi B, Varoni EM et al. Plants of the *Melaleuca* genus as antimicrobial agents: from farm to pharmacy. *Phytotherapy Research* 2017; 31(10): 1475-1494.
- Salehi B, Ayatollahi S, Segura-Carretero A et al. Bioactive chemical compounds in *Eremurus persicus* (Joub. & Spach) Boiss. essential oil and their health implications. *Cellular and Molecular Biology (Noisy le Grand)* 2017; 63(9): 1-7.
- Sharifi-Rad J, Ayatollahi SA, Varoni EM et al. Chemical composition and functional properties of essential oils from *Nepeta schiraziana* Boiss. *Farmacia* 2017; 65(5): 802-812.
- Sharifi-Rad J, Hoseini-Alfatemi S, Sharifi-Rad M, Miri A. Phytochemical screening and antibacterial activity of different parts of the *Prosopis farcta* extracts against methicillin-resistant *Staphylococcus aureus* (MRSA). *Minerva Biotecnologica* 2014; 26(4): 287-293.
- Raeisi S, Ojagh SM, Sharifi-Rad M, Sharifi-Rad J, Quek SY. Evaluation of *Allium paradoxum* (MB) G. Don. and *Eryngium caucasicum* traue. Extracts on the shelf-life and quality of silver carp (*Hypophthalmichthys molitrix*) filets during refrigerated storage. *Journal of Food Safety* 2017; 37(3): e12321.
- Sharifi-Rad M, Varoni EM, Iriti M et al. Carvacrol and Human Health: A Comprehensive Review. *Phytotherapy Research* 2018; doi: 10.1002/ptr.6103.
- Bagheri G, Mirzaei M, Mehrabi R, Sharifi-Rad J. Cytotoxic and Antioxidant Activities of *Alstonia scholaris*, *Alstonia venenata* and *Moringa oleifera* Plants From India. *Jundishapur Journal of Natural Pharmaceutical Products* 2016; 11(3): e31129.
- Sharifi-Rad J, Salehi B, Schnitzler P et al. Susceptibility of herpes simplex virus type 1 to monoterpenes thymol, carvacrol, p-cymene and essential oils of *Sinapis arvensis* L., *Lallemantia royleana* Benth. and *Pulicaria vulgaris* Gaertn. *Cellular and Molecular Biology (Noisy le Grand)* 2017; 63(8): 42-47.
- Sharifi-Rad J, Fallah F, Setzer W, Entezari RH, Sharifi-Rad M. *Tordylium persicum* Boiss. & Hausskn extract: A possible alternative for treatment of pediatric infectious diseases. *Cellular and Molecular Biology (Noisy-le-Grand, France)* 2016; 62(9): 20-26.
- Sharifi-Rad J, Mnayer D, Roointan A et al. Antibacterial activi-

ties of essential oils from Iranian medicinal plants on extended-spectrum β -lactamase-producing *Escherichia coli*. Cellular and Molecular Biology (Noisy-le-Grand, France) 2016; 62(9): 75-82.

28. Hussein S, Usama E-M, Tantawy M, Kawashty S, Saleh N. Phenolics of selected species of Persicaria and Polygonum (Polygonaceae) in Egypt. Arabian Journal of Chemistry 2017; 10(1): 76-81.

29. Lajter I, Zupkó I, Molnár J et al. Antiproliferative activity of Polygonaceae species from the Carpathian Basin against human cancer cell lines. Phytotherapy Research 2013; 27(1): 77-85.

30. Vasas A, Orbán-Gyapai O, Hohmann J. The Genus *Rumex*: Review of traditional uses, phytochemistry and pharmacology. Journal of Ethnopharmacology 2015; 175: 198-228.

31. Zhang Y, Xu H, Chen H, Wang F, Huai H. Diversity of wetland plants used traditionally in China: a literature review. Journal of Ethnobiology and Ethnomedicine 2014; 10(1): 72.

32. Disler M, Ivemeyer S, Hamburger M et al. Ethnoveterinary herbal remedies used by farmers in four north-eastern Swiss cantons (St. Gallen, Thurgau, Appenzell Innerrhoden and Appenzell Auserrhoden). Journal of Ethnobiology and Ethnomedicine 2014; 10(1): 32.

33. Vogl CR, Vogl-Lukasser B, Walkenhorst M. Local knowledge held by farmers in Eastern Tyrol (Austria) about the use of plants to maintain and improve animal health and welfare. Journal of Ethnobiology and Ethnomedicine 2016; 12(1): 40.

34. Bartha SG, Quave CL, Balogh L, Papp N. Ethnoveterinary practices of Covasna County, Transylvania, Romania. Journal of Ethnobiology and Ethnomedicine 2015; 11(1): 35.

35. Cho EJ, Um SI, Han JH et al. The cytoprotective effect of *Rumex aquaticus* Herba extract against hydrogen peroxide-induced oxidative stress in AGS cells. Archives of Pharmacal Research 2016; 39(12): 1739-1747.

36. Shim K-S, Lee B, Ma JY. Water extract of *Rumex crispus* prevents bone loss by inhibiting osteoclastogenesis and inducing osteoblast mineralization. BMC Complementary and Alternative Medicine 2017; 17(1): 483.

37. Wei-Jun F, Jian-Jun T, Hui W et al. In vivo and in vitro anti-sepsis effects of physcion 8-O- β -glucopyranoside extracted from *Rumex japonicus*. Chinese Journal of Natural Medicines 2017; 15(7): 534-539.

38. Hromádková Z, Hirsch J, Ebringerová A. Chemical evaluation of fallopia species leaves and antioxidant properties of their non-cellulosic polysaccharides. Chemical Papers 2010; 64(5): 663-672.

39. Lee S-S, Kim D-H, Yim D-S, Lee S-Y. Anti-inflammatory, analgesic and hepatoprotective effect of semen of *Rumex crispus*. Korean Journal of Pharmacognosy 2007.

40. Inoue M, Nishimura H, Li H, Mizutani J. Allelochemicals from *Polygonum sachalinense* Fr. Schm.(Polygonaceae). Journal of Chemical Ecology 1992; 18(10): 1833-1840.

41. Kang Y, Wang Z, Li J, Liu L. Isolation and identification of two anthraquinones from *Rumex gmelini* Turcz. China Journal of Chinese Materia Medica 1996; 21(12): 741-742.

42. Demirezer LÖ, Kuruüzüm-Uz A, Bergere I, Schiewe H-J, Zeeck A. The structures of antioxidant and cytotoxic agents from natural source: anthraquinones and tannins from roots of *Rumex patientia*. Phytochemistry 2001; 58(8): 1213-1217.

43. Kim J-M, Lee Y-M, Jang D-S, Kim J-S. Constituents of the stems of *Rumex japonicus* with advanced glycation end products (AGEs) and rat lens aldose reductase (RLAR) inhibitory activity. Journal of Applied Biological Chemistry 2006; 49(1): 24-27.

44. Bařkan S, Daut-Özdemir A, Günaydın K, Erim FB. Analysis of anthraquinones in *Rumex crispus* by micellar electrokinetic chromatography. Talanta 2007; 71(2): 747-750.

45. Kim J-M, Jang D-S, Lee Y-M, Lee G-Y, Kim J-S. Constituents of the Fruits of *Rumex japonicus* with Inhibitory Activity on Aldose

Reductase. Journal of Applied Biological Chemistry 2008; 51(1): 13-16.

46. Wang ZY, Zhao HP, Zuo YM, Wang ZQ, Tang XM. Two new C-glucoside oxantrones from *Rumex gmelini*. Chinese Chemical Letters 2009; 20(7): 839-841.

47. Hawas UW, Ahmed EF, Abdelkader AF, Taie HAA. Biological activity of flavonol glycosides from *Rumex dentatus* plant, an Egyptian xerophyte. Journal of Medicinal Plants Research 2011; 5(17): 4239-4243.

48. Jo G, Shin SY, Lee Y et al. A Compound isolated from *Rumex japonicus* induces early growth response gene-1 expression. Journal of the Korean Society for Applied Biological Chemistry 2011; 54(4): 637-643.

49. Ahmed D, Mughal QM, Younas S, Ikram M. Study of phenolic content and urease and alpha-amylase inhibitory activities of methanolic extract of *Rumex acetosella* roots and its sub-fractions in different solvents. Pakistan Journal of Pharmaceutical Sciences 2013; 26(3): 553-559.

50. Ross K, Amanor-Boadu V. System Dynamic Approach to Assessing New Product Introduction: The Case of Functional Foods in the United States. Paper presented at: 2006 Annual meeting, July 23-26, Long Beach, CA, 2006.

51. Cencic A, Chingwaru W. The role of functional foods, nutraceuticals, and food supplements in intestinal health. Nutrients 2010; 2(6): 611-625.

52. Roberfroid MB. Functional foods: concepts and application to inulin and oligofructose. British Journal of Nutrition 2002; 87(S2): S139-S143.

53. Jimoh F, Adedapo A, Aliero A, Afolayan A. Polyphenolic Contents and Biological Activities of *Rumex ecklonianus*. Pharmaceutical Biology 2008; 46(5): 333-340.

54. Martirosyan DM. Functional Foods for Cardiovascular Diseases: Functional Foods Can Help Reduce the Risks of Cardiovascular Diseases: D&A Inc.; 2005.

55. Liang H-X, Dai H-Q, Fu H-A et al. Bioactive compounds from *Rumex* plants. Phytochemistry Letters 2010; 3(4): 181-184.

56. Abbasi AM, Khan SM, Ahmad M, Khan MA, Quave CL, Pieroni A. Botanical ethnoveterinary therapies in three districts of the Lesser Himalayas of Pakistan. Journal of Ethnobiology and Ethnomedicine 2013; 9(1): 84.

57. Hussain SA, Panjagari NR, Singh R, Patil G. Potential herbs and herbal nutraceuticals: food applications and their interactions with food components. Critical Reviews in Food Science and Nutrition 2015; 55(1): 94-122.

58. Dogan Y, Nedelcheva A, Łuczaj u et al. Of the importance of a leaf: the ethnobotany of sarma in Turkey and the Balkans. Journal of Ethnobiology and Ethnomedicine 2015; 11(1): 26.

59. Łuczaj u, Köhler P, Pirożnikow E, Graniszewska M, Pieroni A, Gervasi T. Wild edible plants of Belarus: from Rostafiński's questionnaire of 1883 to the present. Journal of Ethnobiology and Ethnomedicine 2013; 9(1): 21.

60. Ali-Shtayeh MS, Jamous RM, Al-Shafie JH et al. Traditional knowledge of wild edible plants used in Palestine (Northern West Bank): a comparative study. Journal of Ethnobiology and Ethnomedicine 2008; 4(1): 13.

61. Powell B, Ouarghidi A, Johns T, Tattou MI, Eyzaguirre P. Wild leafy vegetable use and knowledge across multiple sites in Morocco: a case study for transmission of local knowledge? Journal of Ethnobiology and Ethnomedicine 2014; 10(1): 34.

62. Łuczaj u, Konèia MZ, Milièevia T, Dolina K, Pandža M. Wild vegetable mixes sold in the markets of Dalmatia (southern Croatia). Journal of Ethnobiology and Ethnomedicine 2013; 9(1): 2.

63. Kasper-Pakosz R, Pietras M, Łuczaj u. Wild and native plants and mushrooms sold in the open-air markets of south-eastern Po-

- land. *Journal of Ethnobiology and Ethnomedicine* 2016; 12(1): 45.
64. Łuczaj u, Pieroni A, Tardío J et al. Wild food plant use in 21st century Europe: the disappearance of old traditions and the search for new cuisines involving wild edibles. *Acta Societatis Botanicorum Poloniae* 2012; 81(4): 359.
65. Williams M, Pehu E, Ragasa C. Functional foods: opportunities and challenges for developing countries. 2006.
66. Ozturk S, Ozturk A. Antibacterial Activity of Aqueous and Methanol Extracts of *Rumex alpinus*. and *Rumex caucasicus*. *Pharmaceutical Biology* 2007; 45(2): 83-87.
67. Bauch H-J, Labadie RP, Leistner E. Biosynthesis of nepodin (2-acetyl-3-methylnaphthalene-1, 8-diol) in *Rumex alpinus* L. *Journal of the Chemical Society, Perkin Transactions 1* 1975; (7): 689-692.
68. Grieve M. Botanical. com: A Modern Herbal. Online document at: <http://botanical.com> Accessed June 2013; 16.
69. Global Invasive Species Database. Species profile: *Rumex obtusifolius*. 2017.
70. Ibáñez-Calero SL, Jullian V, Sauvain M. A new anthraquinone isolated from *Rumex obtusifolius*. *Revista Boliviana de Química* 2009; 26(2): 49-56.
71. Orbán-Gyapai O, Raghavan A, Vasas A, Forgó P, Shah ZA, Hohmann J. Flavonoid-glycosides from *Rumex aquaticus* with neuroprotective activity. *Planta Medica* 2014; 80(16): P11110.
72. Orbán-Gyapai O, Liktör-Busa E, Kúsz N et al. Antibacterial screening of *Rumex* species native to the *Carpathian Basin* and bioactivity-guided isolation of compounds from *Rumex aquaticus*. *Fitoterapia* 2017; 118: 101-106.
73. Fan J, Zhang Z. Studies on the chemical constituents of *Rumex crispus*. *Zhong yao cai- Zhongyaocai- Journal of Chinese Medicinal Materials* 2009; 32(12): 1836-1840.
74. Kargioğlu M, Cenkci S, Serteser A et al. An ethnobotanical survey of inner-West Anatolia, Turkey. *Human Ecology* 2008; 36(5): 763-777.
75. Ahmad S, Ullah F, Zeb A, Ayaz M, Ullah F, Sadiq A. Evaluation of *Rumex hastatus* D. Don for cytotoxic potential against HeLa and NIH/3T3 cell lines: chemical characterization of chloroform fraction and identification of bioactive compounds. *BMC Complementary and Alternative Medicine* 2016; 16(1): 308.
76. Jerezano Alberto V, Ríos Saúl A, Tepancal-Gomez E, Salas-Mendoza E, Villanueva L. Some Traditional Medicinal Plants of North Region from Puebla, Mexico: Uses and Potential Pharmacological Activity of *Rumex* spp. *Natural Products Chemistry and Research* 2016; 4(223): 2.
77. Rao K, Sunitha C, Banji D, Shwetha S, Krishna D. Diuretic activity on different extracts and formulation on aerial parts of *Rumex vesicarius* Linn. *Journal of Chemical and Pharmaceutical Research* 2011; 3(6): 400-408.
78. Zhu J, Zhang C, Zhang M, Wang Z. Studies on chemical constituents in roots of *Rumex dentatus*. *Zhongguo Zhong yao za zhi- Zhongguo zhongyao zazhi- China Journal of Chinese Materia Medica* 2006; 31(20): 1691-1693.
79. Batool R, Aziz E, Tan BK-H, Mahmood T. *Rumex dentatus* Inhibits Cell Proliferation, Arrests Cell Cycle, and Induces Apoptosis in MDA-MB-231 Cells through Suppression of the NF-κB Pathway. *Frontiers in Pharmacology* 2017; 8: 731.
80. Zhou X, Xuan L, Zhang S. Study on the chemical constituents from *Rumex japonicus* Houtt. *Zhong yao cai- Zhongyaocai- Journal of Chinese Medicinal Materials* 2005; 28(2): 104-105.
81. Xie Q-C, Yang Y-P. Anti-proliferative of physcion 8-O-β-glucopyranoside isolated from *Rumex japonicus* Houtt. on A549 cell lines via inducing apoptosis and cell cycle arrest. *BMC Complementary and Alternative Medicine* 2014; 14(1): 377.
82. Girma B, Yimer G, Makonnen E. Effect of *Rumex abyssinicus* on preneoplastic lesions in dimethylhydrazine induced colon carcinogenesis in rats. *BMC Complementary and Alternative Medicine* 2015; 15(1): 365.
83. Useful Tropical Plant, *Rumex abyssinicus*. 2017.
84. El-Hawary SA, Sokkar NM, Ali ZY, Yehia MM. A profile of bioactive compounds of *Rumex vesicarius* L. *Journal of Food Science* 2011; 76(8).
85. Asha Tukappa N, Londonkar RL. Standardization of extraction process for *Rumex vesicarius* L. *International Journal of Scientific and Engineering Research* 2014; 5(4): 1061-1064.
86. Salehi B, Kumar NVA, Şener B, Sharifi-Rad M, Kılıç M, Mahady GB, Vlasisavljevic S et al. Medicinal Plants Used in the Treatment of Human Immunodeficiency Virus. *International Journal of Molecular Sciences* 2018; 19(5): 1459.