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Diabetes up-regulated collagen IV and laminin α5 genes in mRNA and protein levels in seminiferous tubules of C57BL/6 adult mice

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Abstract: Diabetes is a disease associated with impairment of the male reproductive system that causes complications such as decreased testosterone, the diameter of the seminiferous tubule, libido, and fertility. Extracellular matrix (ECM) molecules are involved in testicular development and spermatogenesis. Laminin and collagen are key proteins in seminiferous tubule basement membrane and play an important role in spermatogenesis. The present study was conducted to investigate the effect of diabetes on collagen IV and laminin α 5 changes in mice testis. In this experimental study, 40 mice (C57BL/6) were divided randomly into 4 groups: 1) Control group: without intervention, 2) Diabetic group: treated mice with 50 mg/kg streptozotocin (STZ), 3) Diabetic + Insulin group: treated mice with STZ and insulin, and 4) Sham group: received citrate buffer. After 35 days, the left testes of all specimens were used for Real-Time PCR while their right testes were applied for immunohistochemical study and Periodic acid–Schiff (PAS) staining. This study showed that gene expression and immunoreactivity of laminin α 5 and collagen IV were significantly increased in diabetic mice compared to other groups (P<0.05). Also, PAS staining showed the thickness of seminiferous tubule basement membrane in the Diabetic group compared to other group increased significantly (p<0.05). In Diabetic + Insulin compared to Diabetic group, gene expression, the intensity of immunoreactivity and thickness of seminiferous tubule basement membrane decreased significantly (P<0.05). Our findings indicated that diabetes causes up-regulation of collagen IV and laminin α 5 in mRNA and protein levels in the seminiferous tubule basement membrane and may cause disorder in spermatogenesis in mice.

Key words: Diabetes; Testis; Basement membrane; Laminin a5; Collagen IV.

Introduction

Infertility is considered as one of the serious social problems among nations. In this regard, a male factor is generally involved in half of the infertility problems (1). Several risk factors can influence the male infertility problem, including some diseases such as diabetes. Diabetes mellitus is one of the most common metabolic diseases that are associated with hyperglycemia. Failure to produce insulin causes type I diabetes, peripheral tissue resistant to insulin, and also decreased insulin secretion from pancreatic beta cells generates type II diabetes (2, 3). Long-term diabetes has adverse effects on the visual system, kidney, the cardiovascular, and nervous system (4).

Studies showed that chronic hyperglycemia increases generation of free radicals, particularly reactive oxygen species (ROS), for all tissues (5-8). The studies have shown that diabetes has a variety of functional and structural effects on the male reproductive system and spermatogenesis (9). Several studies have shown that diabetes causes complications like decreased testosterone level, decreased the diameter of seminiferous tubule and libido, increased sperm abnormalities, and infertility (10, 11). Researchers have shown that diabetes may cause changes in the structure of the extracellular matrix (ECM) (12). It has been shown that ECM molecules are involved in the process of development of the testes and spermatogenesis. ECM is a network of proteins and glycoproteins that provides physical support for cells and tissues and plays a role in regulating cell behavior, differentiation and migration. The major ECM components include laminin, fibronectin, collagen I, IV, and VI, versican, and decorin (13-15). Changes in the composition and structure of ECM may alter cell function in different ways. At the molecular level, these changes may be associated with alteration in the metabolism of different types of collagen, fibronectin, and laminin (16). The basement membrane is a part of the extracellular matrix that underlies the seminiferous tubules. This membrane was prepared using Sertoli cells and Myoid cells (17-22). It has been shown that at approximately the second week of pregnancy laminin and collagen IV and heparan sulfate are involved in organizing the undifferentiated testicular cords and fibronectin presents at this location even earlier than this time (12). Collagen IV is a non-fibrillar collagen that is the most abundant protein in the basement membrane and creates about 50% of the basement membrane. Biosynthesis of collagen IV has a complex process encompassing numerous specific and non-specific enzymes. Laminin is the most abundant non-collagenous proteins and plays a role in cell differentiation, cell migration, and cellular junctions. Laminin is composed of three Alpha, Beta,

and Gamma chains. These chains can connect to each other to form at least 15 different isoforms of laminin. Laminins have important functions such as contributing information of basement membrane and act as ligands for receptors on cells (23, 24). As collagen and laminin are two major components of ECM that involve in forming the undifferentiated testicular cords and cell behavior, moreover ECM molecules are involved in the process of development of the testes and spermatogenesis and considering the increasing prevalence of diabetes and its effects on spermatogenesis as well as the importance of the matrix in male infertility, we investigated the effects of diabetes on collagen IV and laminin α 5 expression in mice testes.

Materials and Methods

Animals

For this experimental study, 40 male *C57BL/6* mice aging 2 months with 20-25 g body weight were purchased from Pasteur Institute, Tehran, Iran. The study was done in accordance with the instructions of the National Institute of Health Guide for the Care and Use of Laboratory Animals (NIH Publications No. 80-23, revised 1978) and approved by Ethical Committee at Mashhad University of Medical Sciences, Mashhad, Iran. The mice were housed in standard animal cages and kept under standard laboratory conditions (12h light and dark cycle, at 21°C temperature, and 50% relative humidity). Animals had free access to mice pelleted chow and drinking water during the experimental period.

Study design

The mice were randomly divided into 4 groups. Each group included 10 animals: 1) Control group: without any intervention, 2) Diabetic group: treated mice with Streptozotocin (STZ), 3) Diabetic + Insulin group: treated mice with STZ and insulin (1.5 U of LANSULIN 70/30), and 4) Sham group: that received citrate buffer. Diabetes in mice was induced by intraperitoneal injection of STZ (Sigma company, USA) at a dose of 50 mg/kg in citrate buffer (freshly dissolved in 0.01 mol/L (pH: 4.5) sodium citrate buffer) for 5 consecutive days (25). Next, 72 h after the last injection, diabetes was confirmed by measuring blood glucose levels of the tail vein with a glucometer (Easy Gluco, Infopia, Korea). Mice with blood glucose levels of 250 mg/dL or higher were considered diabetic. At the end of the experimental period after 35 days (spermatogenesis duration in mice) (26), animals were anesthetized and sacrificed

 Table 1. Sequences of primers used for real-time PCR.

by cervical dislocation and testes were removed from each group. The left testes were stored at -70°C for RNA extraction and the right testes were washed in normal saline and fixed in 10% formalin for immunohistochemical study and Periodic acid-Schiff (PAS) staining.

RNA extraction

Total RNA was isolated from testicular tissue by using Total RNA Extraction Kit (Parstous Company, Iran) in accordance with the instruction of the manufacturer. Briefly, the tissue was homogenized in 750 µl of RL solution and then 150 µl chloroform was added to the solution. Afterward, it was incubated at room temperature for 3 min and centrifuged for 12 min, at 13000 rpm. In the next step, 400 µl of the upper phase was transferred into a tube and an equal volume of 70% ethanol was added to the solution. The tube was centrifuged for 1 min at 4°C and 13000 rpm. Then 700 µl and after that 500 µl PW was added respectively and centrifuged for 2 min at 4°C at 13000 rpm and finally, 50 µl Diethyl pyrocarbonate (DEPC) was added and centrifuged for 1 min at 4°C and at 13000 rpm. The purity and integrity of obtained total RNA were checked by 260/280 nm ratio measurement and 1% agarose gel electrophoresis, respectively. Total RNA was kept at -70°C.

cDNA synthesis

Reverse transcription (RT) reagents were purchased and cDNA was synthesized by using cDNA synthesis Kit (Parstous, Company, Iran) according to the manufacturer's instruction. The total RNA (0.5 μ l) was reverse transcribed with 1 μ l oligo (dt) and 8.5 μ l DEPC water. The solution was incubated at 65°C for 5 min. Next, it was immediately transferred to ice and then added 10 μ l reverse transcriptions (RT) premix (reaction volume =20 μ l). The samples were incubated at 50°C for 60 min and then the transcriptase was inactivated at 70°C for 10 min. cDNA samples were stored at -70°C.

Real-time PCR

To assess gene expression, real-time PCR was performed by using the Applied Biosystems Real-Time PCR Instruments (ABI) and SYBER Green/ROX master mix (Parstous Corporation, Iran) kit). To determine gene expression of *laminin* α 5 and *collagen IV*, real-time PCR method was conducted. All samples were tested in duplicate. The primer sequences are described in Table 1. PCR master mix contains 10 µl SYBR Green, 1 µl of each gene-specific primer (forward and reverse), 7.1 µl of distilled water, 0.4 µl ROX, and 0.5 µl of cDNA (volume of reaction = 20 µl in each tube). The PCR pro-

Gene	primer sequences	product size (bp)	Annealing temperature (°C)
Collagen IV	5'-AAGCTGTAAGCATTCGCGTAGTA-3' (R)	107	58
	5'- ATTCCTTTGTGATGCACACCAG-3' (F)		
Laminin α 5	5'-TACCAACGAAGGGCTGCG- 3'(R)		
	5'-CGTCCCACAGGAATAGGCT- 3' (F)	109	58
GAPDH	5'- CTGTAGCCATATTCATTGTCATACCA-3' (R)	385	58
	5'-AACTCCCATTCTTCCACCTTTG-3' (F)		
GAPDH: Glyceraldehyde 3-phosphate dehydrogenase.			

cedure was as follow: keeping at 94°C for 10 min followed by 35 cycles (95°C for 30sc, 60°C for 1min, and 72°C for 30sc). For internal control, Glyceraldehyde 3-phosphate dehydrogenase (*GAPDH*) (27-29) was used to determine the relative expression quantity of the target genes. Real-time data were calculated by using the ratio formula (Ratio= $2-\Delta\Delta$ Ct), where $\Delta\Delta$ Ct = Δ CT (control sample) - Δ CT (target sample) (30-32).

Immunohistochemistry study

For immunohistochemistry study, the specimens, after fixation, were dehydrated with an ascending ethanol series, cleared with xylene, and embedded in paraffin. The blocks were cut into 5 µm sections. Then, tissue sections were deparaffinized and placed in descending concentration of alcohol and then washed with distilled water. Subsequently, sections were rinsed with phosphate buffered saline (PBS) (pH=7.4) for 15 min (3 times). For antigen retrieval, sections were placed in PBS-Ethylene diamine tetraacetic acid (EDTA) solution in a hot water bath for 30 min. After washing with PBS (15 min), in order to block endogenous peroxides activity, sections were immersed in methanol and 3% H2O2 for 20 min. After three times washing in PBS (15 min), nonspecific bindings of antibodies were blocked with bovine serum albumin (BSA) 1% and goat serum 10% for 30 min. Next, the sections were incubated with laminina5 and collagen IV primary antibodies (diluted with 1:100 and 1:200, respectively) overnight at 4°C and the next day was washed with PBS. The samples were incubated with goat polyclonal secondary antibody (1:400) at 37°C for 2 h. After 15 min washing in PBS (3 times), samples were immersed with 0.03% of 3, 3-diaminobenzidine (DAB) in PBS buffer containing 0.01% H2O2 for 15 min, as chromogen. Following the washing, the sections were counterstained with hematoxylin, cleared in xylene, and dehydrated in ascending ethanol series (70-100%). The samples then were mounted with entellan. After imaging with Olympus BX51 light microscope, the reaction was scored semiquantitatively as the following: weak (+), moderate (++), strong (+++), and highly strong (++++) by two persons (30, 33).

PAS staining

The samples were deparaffinized, hydrated and washed with distilled water for 2 min and placed in periodic acid for 10 min. After washing with distilled water, the samples were stained with Schiff for 30 min and then the slides were rinsed in running water and distilled water for 5 and 1 min, respectively. In the next step, the slides were stained in Harris Hematoxylin for 2 min followed by washing in running water for 3 min. The slides were cleared with xylene and mounted. After imaging with Olympus BX51 light microscope, the reaction was scored semi-quantitatively as weak (+), moderate (++), strong (+++), and highly strong (++++) by two persons (34, 35).

Statistical analysis

All statistical analyses were carried out using the Statistical Package for the Social Sciences, version 16, SPSS Inc. Chicago, Illinois, USA (SPSS software). The results of immunohistochemistry and PAS staining were evaluated by Kruskal-Wallis non-parametric test followed by Mann-Whitney test. Also, the results of real-time PCR were analyzed using ANOVA followed by Tukey test. A p value of <0.05 was considered statistically significant.

Results

Expression of laminin $\alpha 5$ and collagen IV in mice testis

Laminin α 5 mRNA expression on testis tissue was detected by real-time PCR. Statistical analyses (Figures 1 and 2) indicated that the relative expression of laminin $\alpha 5$ gene in the Diabetic group was significantly increased compared to that of the Control group (P<0.001). Similarly, there was a statistically significant increase in the expression of laminin $\alpha 5$ in the Diabetic group compared to the Sham group (P<0.001). There was no significant difference between the Control and the Sham groups. There was a statistically significant decrease in the mRNA expression of laminin $\alpha 5$ in Diabetic + Insulin group compare to the Diabetic group (P<0.001) (Figure 1). In the present study, mRNA expression of collagen IV showed similar results of laminin α 5. A statistically significant increase was observed in the Diabetic group in comparison to the Control and Sham groups (P<0.001). Statistical analyses indicated that mRNA expression of collagen IV in the Diabetic + Insulin group was significantly reduced compared to the



Figure 1. Effect of diabetes on laminin α 5 mRNA expression in mice testis. Data expressed as mean \pm SEM. There was a significant increase in Diabetic group compared to the Control and Sham groups *P<0.05 and a significant decrease in the Diabetic + Insulin groups compare to the Diabetic group #P < 0.05. RQ= Relative quantification.



Figure 2. Effect of diabetes on collagen IV mRNA expression in mice testis. Data expressed as mean \pm SEM. There was a significant increase in Diabetic group compared to the Control and Sham groups *P<0.05 and a significant decrease in the Diabetic + Insulin groups compare to the Diabetic group #P < 0.05. RQ= Relative quantification.

Diabetic group (P<0.001). There was no considerable difference between Sham and Control groups (Figure 2).

Immunoreactivity of laminin $\alpha 5$ and collagen IV in mice testis

This study was conducted to evaluate the intensity of laminin $\alpha 5$ and collagen IV proteins in seminiferous tubule basement membrane according to the intensity of color darkness. Our findings showed that the intensity of laminin $\alpha 5$ immunoreactivity in the Diabetic group increased significantly compared to the Control and Sham groups (P<0.001). As presented in Figure. 3, a significant decrease occurred in Diabetic + Insulin compare to Diabetic groups (P<0.001). The results of this study indicated that collagen IV immunoreactivity was similar to laminin a5 immunoreactivity. A considerable increase was also observed in a Diabetic group, compared to the Control and Sham groups (P<0.001). We also found a significant decrease in Diabetic + Insulin group compared to Diabetic group (P<0.001). No remarkable difference was noticed between Sham and Control groups in the intensity of laminin $\alpha 5$ and collagen IV (Figure. 4).

Results of PAS staining in basement membrane of mice seminiferous tubules

Our findings showed that the thicknesses of semi-



Figure 3. The graph shows the effect of diabetes on immunoreaction of laminin α 5 protein in the basement membrane of the seminiferous tubule. There was a significant increase in Diabetic group compare with Control and Sham groups *P<0.05 and a significant decrease in Diabetic + Insulin group compare to Diabetic group #P < 0.05.The photomicrograph shows the effect of diabetes on immunoreactivity of laminin α 5 protein in the basement membrane of the seminiferous tubule; Photomicrographs show immunoreactivity of laminin α 5 protein in Control (A), Diabetic (B), Diabetic + Insulin (C), and Sham (D) groups; Positive immunoreactions show a different grade of intensity brown color (Arrows); Scale bar = 50 µm.

niferous tubule basement membrane in the Diabetic group compared to the Control and Sham groups were increased significantly (p<0.05). Also, a significant increase was observed (p<0.05) in the Diabetic group compared to Diabetic + Insulin group. There was no significant difference among Control, Sham, and Diabetic + Insulin groups (Figure. 5).

Discussion

This study was designed to evaluate the effects of diabetes on laminin a5 and collagen IV genes in mRNA and protein levels in seminiferous tubules of adult mice. The results of this study revealed a statistically significant difference between diabetic and other groups in laminin a5 and collagen IV genes expression and immunoreactivity. Studies showed that in diabetic patients the level of oxidative stress was high due to excess generation of ROS and reduction in efficiency of antioxidant enzyme defenses. Oxidative stress is an important factor in the impairment of male fertility (5). Previous studies have shown that the ECM of most tissues is not well preserved against oxidative stress. Moreover, it has been shown that diabetes may cause changes in the structure of the ECM and the oxidative stress induced by diabetes can cause a change in the gene expression (36). A previous showed that the ECM molecules are



Figure 4. The graph shows the effect of diabetes on immunoreaction of collagen IV protein in the basement membrane of the seminiferous tubule. There was a significant increase in Diabetic group compare with Control and Sham groups *P<0.05 and a significant decrease in Diabetic + Insulin group compare to Diabetic group #P < 0.05.The photomicrograph shows the effect of diabetes on immunoreactivity of collagen IV protein in the basement membrane of the seminiferous tubule; Photomicrographs show immunoreactivity of laminin α 5 protein in Control (A), Diabetic (B), Diabetic + Insulin (C), and Sham (D) groups; Positive immunoreactions show a different grade of intensity brown color (Arrows); Scale bar = 50 µm.



Figure 5. The graph shows the effect of diabetes on basement membrane of seminiferous tubule by PAS staining. There was a significant increase in the Diabetic group compare with the Control and Sham groups *P<0.05 and a significant decrease in the Diabetic + Insulin group compare to Diabetic group #P < 0.05. The photomicrograph shows the thickness of basement membrane by PAS staining in the Control (A), Diabetic (B), Diabetic + Insulin (C) and Sham (D) groups. Scale bar=100 µm.

involved in the development of the testes and spermatogenesis. Among various components of the basement membrane, collagen IV, laminin, and heparan sulfate proteoglycan have been shown to involve in Germ cell and Sertoli function (12, 17, 19, 37, 38). In some spermatogenesis disorders cases, the lamina propria of seminiferous tubule become thick by an increase in the ECM components with differentiation of Myoid cells to fibroblastic nature, fibrosis in peritubular cells, and interstitium with sporadic Leydig cells, as well as edema in interstitium that prevents normal development of germ cells and subsequently male fertility (19, 39, 40). Previous studies have shown that diabetes causes both degeneration and inflammation in the rat's testes (41) and also causes a reduction in seminiferous tubule diameter, increased thickening of the seminiferous tubule basement membrane, and degenerated germ cells in diabetic rats (3). Our results based on immunohistochemical and real-time PCR techniques showed that laminin a5 and collagen IV immunoreactivity and mRNA expression in the Diabetic group was slightly higher compared to the Control group. Also, PAS staining showed that thickness of seminiferous tubule basement membrane was increased significantly in the Diabetic group compared to other groups. Davis et al. conducted a study on differential expression in the Sertoli cells extracellular matrix. Using immunofluorescence they revealed that the highest expression of laminin and fibronectin occurred during the period of evolution (1-10 days) and then

maintained at a low level while the expression of collagen IV reaches its peak at a later stage (17). The present study demonstrated effects of diabetes on changes in the amount of collagen IV and laminin $\alpha 5$, through evaluating the samples by immunohistochemistry and real-time PCR techniques. Laminin a5 and collagen IV level in the Diabetic group increased while in the Diabetic + Insulin group both laminin $\alpha 5$ and collagen IV levels was statistically significant decrease. In another study, it was demonstrated that diabetes affects the kidney extracellular matrix, they found that the amount of collagen IV decreased while the amount of laminin $\alpha 5$ increased (42). Similarly, the results of our study showed an increase in the laminin $\alpha 5$ rate in the Diabetic group; however, unlike their findings in our study, the amount of collagen IV also increased. In addition, our study showed that in the group that received insulin, both the collagen and laminin $\alpha 5$ were significantly reduced compared to the Diabetic group. Moreover, in the Diabetic + Insulin group compared to the Diabetic group, the thickness of seminiferous tubule basement membrane by using PAS staining was reduced significantly. Because the ECM of most tissues is not well preserved against oxidative stress, an increase occurred in the sensitivity of tissues to proteolytic cleavage and ECM permeability, leading to the altered function of the organ (38, 43). Previous studies have shown that diabetes increases oxidative stress in the testis and hyperglycemia decreases the antioxidant defenses system and reactive oxygen species overproduction (5, 6, 41, 6)44, 45). Moreover, it has been reported that the ECM of most tissues is not well preserved against oxidative stress (38). As that in previous studies, an increase in oxidative stress associated with diabetes was observed. Moreover, it was observed that oxidative stress could lead to some changes in the expression of the gene in the testis tissue. Therefore, oxidative stress is likely to be an effective factor in our study. However, further investigation clearly is warranted.

The results of this study indicated that diabetes causes spermatogenesis disorder in adult mice via upregulation of Collagen IV and Laminin $\alpha 5$ genes in mRNA and protein levels in the seminiferous tubule basement membrane.

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Conflict of interest

The authors confirm that there is no conflict of interest regarding the publication of this article

Author's contributions

Elnaz Khordad, Mohammad Reza Nikravesh , Mehdi Jalali , Alireza Fazel, Mojtaba Sankian, Fatemeh Alipour.

Kh.E.; performed experiments, analysed data, Performed transporter experiments, co-wrote the paper, Provided essential mouse strains, N.Mr.; Supervised the research, co-wrote the paper, Designed experiments,J.M.; Supervised the research, co-wrote the paper, Designed experiments,F.AR.; Consultant, S.M.; Consultant,A.F.; performed experiments,co-wrote the paper.

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