

## Cellular and Molecular Biology

CM B Association

Journal homepage: www.cellmolbiol.org

# Diagnosis and detection of VicK gene in *Streptococcus mutans* isolated from the saliva of patients with diabetic type 2 with tooth decay in the Iraqi population

Susan F.Khadhem Al-Sudani<sup>1</sup>, Laheeb R. Hamad<sup>2</sup>, Fattma A. Ali<sup>3\*</sup>

<sup>1</sup>Lecture in Medical Microbiology, Pathology & Genetic, College of Dentistry, Al-Bayan University, Baghdad, Iraq <sup>2</sup>Lecturer in Microbiology, Department of Pathological Analysis, College of Applied Sciences, Al Fallujah University, Iraq <sup>3</sup>Assistant Professor in Medical Microbiology, Medical Microbiology Department, College of Health Sciences, Hawler Medical University, Kurdistan Region-Iraq

#### **ARTICLE INFO**

#### ABSTRACT

*Original paper Article history:* Received: August 06, 2021 Accepted: November 29, 2021 Published: December 01, 2021

Keywords: Diabetes mellitus (T2DM), VicK gene, Streptococcus mutans, PCR Sequencing Type 2 diabetes mellitus (T2DM) is gradually becoming more common in Iraq. Salivary changes and proliferation of specific bacterial communities cause oral disease that can adversely affect systemic conditions such as diabetes. Fifty saliva samples were collected from people with T2DM suffering from tooth decay and twenty-five people without T2DM suffering from tooth decay. The periodontal status, the extent of the root surface, and coronal caries were evaluated. Saliva was cultured for investigating Streptococcus mutans. The results showed that patients with type 2 diabetes had significantly more severe Periodontitis and a higher prevalence and magnitude of bacterial caries. Diabetic subjects had higher levels of Hemoglobin A1c (HbA1c) and Random Blood Sugar (R.B.S.). The S. mutans diagnosis by PCR for Sanger Sequencing technique by using VicK gene sequences (1300bp). The PCR products of the isolate were submitted to Macrogen Company for sequencing. Selected seven isolates as new isolates registered in global gene bank as locally S. Mutans isolates in Bagdad city/Iraq and their accepted accession numbers include LOCUS MT603520, MT603521, MT603522, MT603523, MT603524, MT603525, and MT603526 of nucleotide sequence. The VicK genes isolates' phylogenetic trees revealed a genotype that was closely connected to other isolates in GenBank. Furthermore, gene sequencing demonstrated a success rate of 99 percent. resemblance to other isolates in the GenBank database The likelihood of a link between S. Mutans and dental carries was determined by these findings.

DOI: http://dx.doi.org/10.14715/cmb/2021.67.4.25 Copyright: © 2021 by the C.M.B. Association. All rights reserved.

### Introduction

Diabetes mellitus (D.M.) is a group of metabolic diseases with hyperglycemia caused by insulin action and/or insulin secretion defects (1). Type 1 diabetes, type 2 diabetes, and gestational diabetes are the three main types of diabetes (2). Type 2 diabetes is the most common. Chronic hyperglycemia is linked with longterm tissue damage, which results in dysfunction and failure of different organs (3,4) and an increased risk of oral diseases, inclusively periodontal disease (5,6), dental caries (7), and xerostomia (8). Diabetes mellitus causes structural and functional changes in susceptible tissues, leading to certain complications in type 1 D.M. and type 2 D.M. Individuals with no functional metabolic abnormalities, insulin deficiency, and hyperglycemia are generally at low risk for developing these complications. Oral health depends on the stability of microbial communities, and oral disease occurs when pathogenic species outnumber the normal flora (9). Pathogenic microorganisms in the oral cavity are frequently linked to two primary diseases: dental caries and periodontal disease (10).

Non-enzymatic glycosylation (11), altered hemodynamics (12), and genetic factors are involved in the development of D.M. (13-16). However, it is clear which factors or combinations not of mechanisms are directly responsible for complications in the target tissues. It is also not clear whether a different type of mechanism is operating in different tissues. However, it is known from prior studies that not all people with diabetes mellitus experience these complications; also, there is a variation in the developmental severity rate and of these complications.

Many studies have supported the idea that patients with diabetes mellitus are at higher risk of developing periodontitis. In other words, periodontal disorder is considered an additional complication of diabetes (17). Periodontal disease is a widespread chronic inflammatory disorder in which the supporting structures of teeth are damaged (the periodontal ligament and alveolar bone). It is widespread (severe periodontitis affects 10-15% of adults) and has many detrimental effects on life quality. The relationship between the degree of hyperglycemia and the severity Periodontitis is evident. The underlying of mechanisms of the association between these two conditions are not entirely known but include aspects of immune functioning, cytokine biology, and neutrophil development. Evidence is emerging to support the presence of a two-way relationship between diabetes and periodontitis, as diabetes enhances the risk of periodontitis and, on the other hand, glycemic regulation is adversely affected by periodontal inflammation. The prevalence of macroalbuminuria and end-stage renal disease is two to three times higher in diabetics with severe periodontitis than in diabetics without severe periodontitis. Various researches had conducted to find out the relationship between glycemic control and Periodontitis. Findings suggest that poor glycemic control is associated with an increased risk of periodontitis (18–25). However, multiple studies have reported no association between glycemic control and periodontitis (17,26-32). Most of these researches usually investigated type 1 diabetes mellitus or do not determine the type of DM. Many of them failed to estimate the glycemic control-related risk of periodontal disorder as they are cross-sectional studies. Limited studies used multivariate analysis, but no study has used teeth or oral health as analysis components for associated observation in their statistical analysis. In spite of very little research attention, researchers investigated the possible relation of dental caries with diabetes (33,34). The outcomes confirmed that there is no difference between people with diabetes and healthy individuals in Finland in the incidence of dental caries, while another study indicated that diabetes was a significant risk agent for root caries (7). Although the causes for the increased incidence of diabetic dental caries remain unclear (34), many oral pathogens, like *Streptococcus mutans* (S. mutans) and lactobacilli in the saliva, may be associated with this. However, in two separate studies, reports on S. mutans and lactobacilli in the saliva of diabetic and non-diabetic individuals were not substantially different (7). Vic-genes regulate the expression of multiple genes associated with virulence (35). Furthermore, VicK inactivation results in a decreased level of lactic acid and an increase in acid tolerance for S. Mutans (36). A VicK knockout mutant was found to be more responsive to H2O2 than the wild-type mutant (37).

The aim of this study was the evaluation the effects of Diabetic Mellitus on dental caries and specifically Streptococcus mutans and investigation of their phylogenetic tree.

### Materials and methods Ethical consideration

The bacterial strains used in this research were extracted from clinical routine specimens, and patients were given verbal consent. This study has been accepted by the College of Health Sciences / Hawler Medical University Scientific and Research Ethics Committee

## Sample Collection

A total of fifty patients with diabetes mellitus and tooth decay had classified into two groups (group1: patients with T2DM for ten years, and group 2: patients with T2DM for 15 years), and also, as a control group, twenty-five individuals were attended in this study. They were examined in National Diabetes Center, Mustansirya University, Baghdad, Iraq, between September 2019 and January 2020. The Hemoglobin A1c (HbA1c) and Random Blood Sugar (R.B.S.) were evaluated by BioSystems Analyzer (A15 Automated Analyzer). Careful history was obtained from both groups according to а questionnaire that covers all information, including name, age, family history, sex, smoking, drinking alcohol and any other diseases after diagnosis by medical staff.

### Culture of sample and Morphology of bacteria

Saliva samples were taken from carious lesions of fasting patients who had their usual examination in the morning using the ends of sterile wooden toothpicks. Before collecting the samples, they were told not to brush their teeth for at least an hour. To avoid dilution of samples, the individuals were instructed to rinse their mouth with distilled water and wait at least 5 minutes. The toothpicks were cut off and dipped in 1 mL of sterile phosphate-buffered saline (PBS) (HiMedia, India) before being stored at 4 °C. To scatter the plaque and achieve a homogenous suspension, saliva samples were vortexed for one minute. The samples were diluted 100-fold in 1x sterile PBS and plated on Mitis Salivarius Bacitracin (M.S.B.) agar. Salivarius agar mitis (HiMedia, India) was used to make the M.S.B. agar, which was supplemented with 15% sucrose, 1% agar, 0.0001% potassium tellurite solution, and 0.2 units/ml bacitracin (HiMedia, India). The plates were incubated anaerobically at 37°C for 48 h. The were recognized based colony colonies on morphology after the incubation phase. Each sample plate's typical colonies were transferred to brain-heart infusion (B.H.I.) broth (HiMedia, India) and cultured for 18 hours at 37°C. The broth cultures were streaked on M.S.B. agar and anaerobically incubated at 37°C for 48 hours after the incubation period. The overnight bacterial cultures were kept at 20°C in a stock of 80 percent glycerol (38).

## 16S rRNA PCR for *Streptococcus mutans* identification

The identity of Streptococcus mutans was verified via po¬lymerase chain reaction (PCR). All primers for the detection of 16SrRNA genes were designed using the NCBI primer designing tool. The primers were synthesized and provided by Bioneer Company, Korea (F: GTTTACGGCGTGGACTACCA and R: CCACACTGGGACTGAGACAC). The final volume of the PCR reaction was 25  $\mu$ L using 12.5  $\mu$ L of 2x Hot Start Taq Master Mix, 1  $\mu$ L of the DNA template, 1  $\mu$ L of each primer (20 pmol) and 9.5  $\mu$ L of ddH2O. DNA amplification was done in a thermocycler PCR. Amplified products were subjected to electrophoresis using 1.2% gel agarose (GeNet Bio, Korea) (Figure 1).

## Molecular diagnosis of VicK gene using PCR technique

Total DNA of Bacteria isolates were extracted using DNA Extraction Mini Kit (17045/ Intron biotechnology/Korea). The VicK gene was amplified using the primer F (5'-CGGGATCCATGACTAATGTGTTTGAATCAAGT C -3') and R (5'-CCGCTCGAGTCATGATTCGTCTTCATCTTCTTC C -3') (36). The PCR amplification was done in a total volume of 25 $\mu$ l containing 1.5 $\mu$ l DNA, 5 $\mu$ l Taq PCR Pre Mix, 1 $\mu$ l of each 16.5 pmol primers which is applied in a tube with a total volume of 25 $\mu$ l then nuclease-free. Thermo cycling conditions were as seen in Table 1.

devices			
Steps	Temperature	Time	No. of Cycles
Denaturation 1	95°C	5min	1
Denaturation 2	95°C	45sec	
Annealing	48°C	45sec	35
Extension 1	72°C	45sec	55
Extension 2	72°C	7min	1

 Table 1. PCR program that was applied in the thermocycler devices

### Sequence of gene

VicK gene sequencing was conducted at Macrogen Company using the ABI 3730xl genetic analyzer (Applied Biosystems, U.S.A). Homology searches were carried out online at the National Center for Biotechnology Information (NCBI) at (http://www.ncbi.nlm.nih.gov) and Bio Edit utilizing the Basic Local Alignment Search Tool (BLAST) program. The outcomes were compared with information accessible online at the NCBI from the ExPASY program released by Gene Bank.

### **Results and discussion**

This study detected VicK gene in samples collected from the saliva of patients with T2DM. PCR products showed that 3 (30%) out of 10 positive samples of S. Mutans in G1 group were positive for VicK gene and 5 (38.4%) out of 13 isolates in group 2 were positive for this gene as seen in Table 2.

Table 2.	Distribution	of positive	Streptococcus	mutans
isolates.				

Isolates	No. of isolate (%)	No. of Morphology of S. Mutans (%)	No. of positive VicK gene product (%)
G1 group	25 (33.3%)	10 (40%)	3 (30%)
G2 group	25 (33.3%)	13 (52%)	5 (38.4%)
Control group	25 (33.3%)	3 (12%)	0

All isolates in our study have been investigated for detection of S. mutans by 16S rRNA genes (Figure 1) with a product size of 120 bp. 16S rRNA-based PCR assays quickly, easily and reliably identify Streptococcus mutans and distinguish them from other phylogenetically related Pseudomonas spp. Assays have a sensitivity and accuracy of 100 % of intended targets. Amplification of 16S rRNA from 22 isolates was performed to confirm bacterial identification. Primers for the conserved region of 16S rRNA were designed and used for PCR amplification of DNA of S. mutans isolates. Then PCR products were separated on agarose gel (Figure 1). The result demonstrated that 10(33%) of S. mutans had 16S rRNA gene band with 120 bp. The result shows that patients with T2DM have significantly higher Hemoglobin A1c (mmol/mI) levels compared to the control group (Figure 2). The results, however, revealed no significant differences between T2DM groups (group1: patients with type 2 diabetes for up to 10 years, and group2: patients with type 2 diabetes for 15 years or more).

The values of serum Random Blood Sugar (mg/dl), which are illustrated in Figure 3, revealed a significantly higher level in the patient's groups compared to the control group. The polymerase chain reaction diagnostic techniques are rapid, easy, inexpensive protocols, becoming the most commonly utilized molecular genetics method for detecting essential genes and identifying the bacteria. The results of PCR amplification of VicK gene (1300bp) were performed and results are shown in Figure 4.



**Figure 1.** Agarose gel electrophoresis of PCR products after amplification of the 16S rRNA gene (120 bp). Lane M: 100 bp DNA ladder. Lane 1: Negative control. Lane 4, 7 and 11: Negative control. Lanes 2,3,5,,6,,8,9,10,13,14,15: Positives for *Streptococcus mutans* 

The phylogenetic tree in Figure 5 was generated with the Molecular Evolutionary Genetics Analysis (MEGA) software version 6.0. For phylogenetic analysis, a neighbor-joining tree was built (Figure 5).





Figure 2. Levels of HbA1C (mmol/mI) of T2DM patients and control subjects



Figure 3. Levels of serum R.B.S (mg/dl) of T2DM patients and control healthy subjects



**Figure 4.** Agarose gel electrophoresis for VicK *gene* (1300bp). Bands were fractionated by electrophoresis on 2% agarose gel (2 h., 5V/cm, 1X T.B.E.) and visualized under U.V. light after staining with a red stain. Lane M: 100bp ladder, Lane 2, 3 and 11: Positive PCR product of G1, Lane 16, 17, 24, 27 and 30: Positive PCR product of G2



Figure 5. Neighbor-joining tree *Streptococcus mutans* of VicK gene

The results of alignments demonstrated that the *Streptococcus mutans* of Iraq and other global strains show partial sequence similarity in translating specific regions of the VicK gene. Hierarchical cluster analysis determines the clusters including *Streptococcus mutans* Iraq isolates of 2 and 3 with the identical percentage of 95%, isolates of 1,4,5,6,7 and 8 with the identical percentage of 97%, and 98 % it is close to Iraq (ID: MN427434) the with the identical percent of 98%.

Distance matrices and the results of BLAST which compared the *VicK* gene isolated from the *Streptococcus mutans* in this study other global isolates are presented in Figure 6.

The sequencing of the VicK gene was performed by the Macrogen Company. The findings were compared with data obtained from the ExPASY system published by Gene Bank available online at the NCBI. The results of *S. Mutans* positive isolates showed that more variation in isolates was seen in group2 which showed 11 variations including five Transversion and six Transition. On the other hand, group 1 showed eight variations including two Transversion and six Transition which 98% identified with the standard of Gene Bank as was shown in Table (3).

								1		2	3	4		5	6
1. 1 MT6	603520.1 Strepts	0000	cus	s mutar	is strai	n FaSu-	I IRAQ								
2. 2 MT6	603521.1 Strepts	0000	cus	s mutar	is strai	n FaSu-:	11 IRAQ	0.	001						
3. 3 MT6	603522. 1 Strepts	0000	cus	s mutar	is strai	n FaSu-S	50 IRAQ	0.	004	0.003					
4. 4 MT(	603523. 1 Strepts	0000	cus	s mutar	is strai	n FaSu-{	89 IRAQ	0.	002	0.001	0.004				
5. 5 MT6	603524. 1 Strepts	0000	cus	s mutar	is strai	n FaSu-	19 IRAQ	0.	002	0.001	0.004	0.00	2		
6. 6 MT6	603525.1 Strepts	0000	:005	s mutar	is strai	n FaSu-	16 IRAQ	0.	002	0.001	0.004	0.00	2	0.000	
7. 7 MT(	603526.1 Strepts	0000	cus	s mutar	is strai	n FaSu-	161 IRAC	2 0.	002	0.001	0.004	0.00	2	0.000	0.000
Spec	ies/Abbrv														
1. 1	MT603520	.1	S	Strer	toc	occus	muta	ans	st:	rain	FaSu-	1 IF	0AS		_
2. 2	MT603521	.1	S	Strer	toc	occus	muta	ans	st	rain	FaSu-	11 I	RA	)	
3. 3	MT603522	.1	5	Strer	toc	occus	mista	ans	at	rain	FaSu-	50 T	RA	~ )	
4 4	MT603523	1	9	Strer	toc	occus	milta	ang	et.	rain	FaSu-	89 T	RA	× D	_
	MT602524	1	0	Strop	1000	00000	muto		at	nain	Facu	10 I	DN/	× n	
5. 5	MTCO2524	• +	 	orret Sener		occus	muut	1115	36.	Lain	FaSu-	15 1	INH.	~	
0. 0	MI603525	•1	2	otrep	COCO	occus	muta	ans	St:	rain	rasu-	10 1	RA	4	_
7. 7	M1603526	.1	5	strep	toc	occus	muta	ans	st	rain	rasu-	161	IR	AQ	
<ol> <li>Classical Control (Control (Contro) (Contro) (Contro) (Contro) (Contro)</li></ol>	I I I I I I I I I I I I I I I I I I I	AATAA AATAA GATTI GATTI GATTI GATTI GATTI GATTI GATTI TATIA TIATIA TIATIA TIATIA TIATIA	ATAA ATAA TAT TAT TAT TAT TAT TAT TAT T	ALCAAU ALCAAU CAGAAGTI CAGAGTI CAGAGT	ARGUART TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT TTTCCGT	TRARIECT TRARECT TRARECTA TTARECTA TTAR	IGAAATTTI IGAAATTTI	ICTITG GGCRCA GGCRCA GGCRCA GGCRCA GGCRCA GGCRCA GGCRCA GGCRCA AGGCRTT AGGCRTT AGGCRTT AGGCRTT	ATTAC ATTAC GOAGA	UBECCATTA TGBECCATTA RARACCEST RARAC	TACTGATAAA TACTGATAAA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA TGACTAGTA CATATTTGAA CATATTTTGAA CATATTTTGAA CATATTTTGAA	ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC ITTTAIC	TTATA TTATA TTATA TTATA TTATA TTATA TTATA TTATA TTATA STTATA STTATA STTATA	IGATARCI IGATARCI TGACAGA TGACAGA TGACAGA TGACAGA TGACAGA TGACAGA TGACAGA TGACAGA TTACAAT TTACAAT TTACAAT	ICGACITEJ ICGIGACITEJ ICGIGITITI ICGIGITI ICGIGITITI ICGIGITITI ICGIGITITI ICGIGITITI ICGIGITI
Sec 1. 1 CGGAA 2. 2 CGGAA 3. 3 CGGAA 3. 3 CGGAA 5. 5 CGGAA 5. 5 CGGAA 5. 5 CGGAA 7. 7 CGGAA 5. 5 CGGAA 7. 7 CGGAA 5. 5 CGGAA 7. 7 CGGAA 5. 5 CGGAA 5		TGAATA TGAATA TGAATA TGAATA TGAATA TGAATA TGAATA TGAATA TGAATA TGAATA AATGT AATGT AATGT AATGT AATGT AATGT	ATGA ATGA ATGA ATGA ATGA ATGA TCAG TCAG	ATGAATTTA ATGAATTTA ATGAATTTA ATGAATTTA ATGAATTTA ATGAATTTA ATGAATTTA GTCATGAA GTCATGAA GTCATGAA GTCATGAA	ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA ICACIITA	CSCATTCSTT CSCATTCSTT CSCATTCSTT CSCATTCSTT CSCATTCSTT CSCATTCSTT CSCATTCSTT CSCCATTAR CACCATTAR CACCATTAR CACCATTAR	ITECCITAAN ITECCITAAN ITECCITAAN ITECCITAAN ITECCITAAN ITECCITAAN CCICIETAA CCICIETAA CCICIETAA CCICIETAA CCICIETAA CCICIETAA	CCSCCGT CCSCCGT CCSCCGT CCSCCGT CCSCCGT CCSCCGT AGTCCT AGTCCT AGTCCT AGTCCT AGTCCT AGTCCT	GAAAGT GAAAGT GAAAGT GAAAGT GAAAGT GAAAGT GAAAGT ATTTAI ATTTAI ATTTAI ATTTAI ATTTAI	GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGCITIATIT GGGCITIG GAAGCITIG GAAGCITIG GAAGCITIG	CAGGACTAATT CAGGACTAATT CAGGACTAATT CAGGACTAATT CAGGACTAATT CAGGACTAATT CAGGACTAATT CAGGACTGATG GACGATGGTG GACGATGGTG GACGATGGTG GACGATGGTG GACGATGGTG GACGATGGTG	COLLECTER COLLEC	SCATGA SCATGA SCATGA SCATGA SCATGA SCATGA SCATGA SCATGA AGAAT( AGAAT( AGAAT)	receacted receac	ACAGGAAAAA ACAGGAAAAAA ACAGGAAAAA ACAGGAAAAAA ACAGGAAAAAA ACAGGAAAAAA ACAGGAAAAAAAA
See			TURE	GTCATGAR	CTGCGGA	CACCATTAN	CCTCIGIAN	ACIUCI		SERGUIIIG	SACGATGGTG	CCCTGAC	COARL .	Reiseri	

**Figure 6.** Distance matrices and the results of BLAST which compared the *VicK* gene isolated from the *Streptococcus mutans* in this study to other global isolates

No. of sample	Type of substitution	Location	Nucleotide	Sequence ID of gene Bank	Sequence ID of Iraqi Strain
	Transition	136	A>G		
	Transition	704	A>G		
	Transition	877	A>G		
Crown 1	Transition	993	A>G	CD050272 1	MT461609 1
Group1	Transversion	997	C>G	CP050275.1	W11401098.1
	Transition	1001	C>T		
	Transversion	1012	T>A		
	Transition	1029	T>C		
-	Transition	136	A>G		
	Transversion	241	A>T		
Group 2	Transversion	244	A>T	AD012226	
	Transversion	264	T>A	AP012550	
	Transition	704	A>G		
	Transition	877	A>G		
	Transition	993	A>G		
	Transversion	997	C>G		
	Transition	1001	C>T		
	Transversion	1012	T>A		
	Transition	1029	T>C		

Table 3. Represents the type of polymorphism from VicK gene from *Streptococcus mutans* for isolates Group 1 and Group 2.

The current investigation offers information on the connection between diabetes and oral disease. There is a link between bacterial numbers and active caries in individuals with diabetes. The higher frequency of functional dentistry caries in patients with diabetes than non-diabetics patients was supported by previous studies (7).

Diabetes mellitus is a disorder that causes elevated glucose levels in the blood resulting from decreased blood insulin levels. This causes several metabolic defects in the carbohydrate, fat, and protein pathways (39). HbA1c assays measure the amount of haemoglobin glycated in the blood and provide a reliable estimation of blood glucose regulation over the last 1 to 3 months. Glycation gradually appears in about 2 to 3 months and recurs early in its development and remains constant until the regeneration of red blood cells (R.B.C). That's why HbA1c has been used to monitor people with diabetes as an index of long-term glycemic control (40). D.M. also affects the mouth and can lead to dental caries, periodontitis, low salivary level, oral mucosal diseases, and infections such as lichen planus, recurrent aphthous stomatitis and candidiasis. Oral health is vital for a person with diabetes. Epidemiological studies indicate that diabetes is a major risk factor for periodontitis and that if glycaemic regulation is low, the risk of periodontitis is higher. People with poorly controlled diabetes (who are also at great risk of other macrovascular and microvascular complications) are at increased risk of periodontitis and loss of alveolar bone (41,42). Given the expected rises in the prevalence of diabetes over the next few decades, the previously observed declines in the majority of Periodontitis (associated with less smoking and improved oral health behavior in recent years) are likely to be reversed as a consequence of a significant rise in the number of people with diabetes (43). It is possible that diabetes

management (i.e., improving glycaemic control) would decrease the risk and severity of Periodontitis. Evidence indicates that periodontal inflammation resolution may improve metabolic control (with approximately 0.4 percent reduction of HbA1c reported), although massive, multi-center, randomized controlled trials are required to validate these findings further.

The main symptoms of periodontal disease are gingival bleeding, and the saliva's decreased flow rate is a vital risk agent for oral candidiasis (44). Dental carries and D.M. have a complicated relationship. Diets that restrict carbohydrate intake are commonly prescribed for children with T1DM., cariogenic foods, T2DM is primarily connected with obesity and a highcalorie, carbohydrate-rich diet in children and adults. There is no clear trend in the literature concerning the connection between dental caries and diabetes (45).

A recent study in Saudi Arabia has shown a high prevalence of oral bacterial in subgingival pockets of T2DM patients as compared to normal subjects (46). The streptococcus mutans were selected based on previous studies. For example, a study that tested different patients with dental caries accompanied by diabetes mellitus showed significantly high numbers of streptococcus mutans compared to other bacterial infections (47). Furthermore, another study demonstrated that patients who suffered from dental plaque and were positive for S. Mutans showed higher caries incidence (48). S. Mutans from dental carries in Iraq was investigated previously by many studies and had been shown various frequencies. A study was done in the Thi-Qar governorate in Iraq and the data demonstrated 33 (41%) S.mutans isolates were isolated from 80 dental plaque individuals aged between 7 to 15 years (49). Mahdi (2015) in Kufa, Iraq, reported that the ratio of S.mutans in dental caries patients is 40%. In a study conducted in Saudi Arabia, Almusawi et al.(2020) discovered that a considerable number of T2DM patients(78%) had high counts of streptococcus mutans in their saliva (105 CFU/ml) and significant relationships between streptococcus mutans load and diabetes., saliva flow rate, saliva buffering capacity, and glycemic control (50). Streptococcus mutans thrived in the saliva of T2DM patients because of hyposalivation, increased salivary glucose, and poor glycemic control. In this study, we obtained a ratio of 40% of S.mutans

isolates in the first group and 52% in the second group. The PCR amplification has confirmed the results and showed 33.3% in both groups, which is lower than previously reported in studies.

Identification of *S. mutans* isolates using 16S rRNA is more accurate than bacteriological and biochemical assays. Rampini *et al* (2011) demonstrate that 16S rRNA gene PCR which was sensitive and specific to use for diagnosis of culture-negative bacterial infections is also useful for identification of bacterial pathogens in patients pretreated with antibiotics (51).

One *S. mutans* bacterial isolate from Iraq has been registered in the global gene bank after examination of the VicK gene sequence. Locally *S. mutans* isolates in Baghdad city which is not similar to global isolates can be considered as new isolates. Therefore, selected isolates were registered as new isolates in the global gene bank and their accepted accession numbers include LOCUS MT603520, MT603521, MT603522, MT603523, MT603524, MT603525, and MT603526 of the nucleotide sequence. The phylogenetic trees for the VicK gene, which encodes the *S. mutans*, revealed a genotype that was closely connected to others in Gen Bank. Gene sequencing found 99 percent similarity with other isolates in GenBank, similar to the *S.mutans* gene.

In this study, we sequenced the vicK gene of S. Mutans strains isolated from two groups of patients with dental caries to analyze the effects of vicK polymorphisms. VicK gene is responsible for regulating the expression of multiple virulenceassociated genes, which affect polysaccharide and synthesis adhesion. Additionally, vicK inactivation produces a reduced level of lactic acid. It improves acid tolerance S. Mutans, so any polymorphism account in this gene can affect the surveillance and virulence of S. Mutans. The capacity of S.Mutans to consume glucose as a result of carbohydrate metabolism to generate lactate is a crucial virulence factor associated with this pathogen. It was shown that the inactive Vick gene could reduce acid production, which affects bacterial surveillance.

In this study, both groups' sequencing results showed variations in the vicK gene. The first group showed 11 variations, while the second group showed eight variations. These results indicated the probability of the relationship between *S. Mutans* and dental carriers. Clinical isolates of mutans and the vicK genes C470 T missense mutation may be linked to the caries experience with S. mutans. The phylogenetic analysis was done for geographic genetic distance determination. The analyzes of phylogenic distance occurred for partial-length of vicK. The results provided a clear picture of genetic distance. The isolates 2 and 3 showed the same distance, and they differ from the other Iraqi isolate published previously (I.D.; MN427434) by only 3%. We can suggest from the results of this study that variation is dependent on how antibiotic treatment can be effective.

### Conclusion

These results indicated High prevalence of oral bacterial *S. Mutans* in Diabetic patients and the probability of the relationship between *S. Mutans* and dental carries and clinical isolates of mutans and the vicK genes missense mutation may be linked to the caries experience with *S. mutans*.

### Acknowledgments

None

### **Interest conflict**

The authors declare no conflict of interest.

### References

1. Kahn CR, White M. The insulin receptor and the molecular mechanism of insulin action. J Clin Invest 1988, 82:1151–6.

2. Association AD. Diagnosis and Classification of Diabetes Mellitus. Diabetes Care 2010, 33:S62 LP-S69.

3. Brown WV. Microvascular complications of diabetes mellitus: renal protection accompanies cardiovascular protection. Am J Cardiol 2008, 102:10L-13L.

4. Bodiga VL, Eda SR, Bodiga S. Advanced glycation end products: role in pathology of diabetic cardiomyopathy. Heart Fail Rev 2014, 19:49–63.

5. Azeez S, Jafar, S., Aziziaram, Z., Fang, L., Mawlood, A., Ercisli, M. Insulin-producing cells from bone marrow stem cells versus injectable insulin for the treatment of rats with type I diabetes. Cell Mol Biomed Rep 2021, 1(1): 42-51.

6. Kampoo K, Teanpaisan R, Ledder RG, McBain AJ. Oral bacterial communities in individuals with

type 2 diabetes who live in southern Thailand. Appl Environ Microbiol 2014, 80:662–71.

7. Hintao J, Teanpaisan R, Chongsuvivatwong V, Dahlen G, Rattarasarn C. Root surface and coronal caries in adults with type 2 diabetes mellitus. Community Dent Oral Epidemiol 2007, 35:302–9.

8. Soell M, Hassan M, Miliauskaite A, Haikel Y, Selimovic D. The oral cavity of elderly patients in diabetes. Diabetes Metab 2007, 33:S10–8.

9. Jakubovics NS, Palmer RJ. Oral microbial ecology: Current research and new perspectives 2013.

10. Li X, Kolltveit KM, Tronstad L, Olsen I. Systemic diseases caused by oral infection. Clin Microbiol Rev 2000, 13:547–58.

11. BROWNLEE M. Nonenzymatic Glycosylation and the Pathogenesis of Diabetic Complications. Ann Intern Med 1984, 101:527.

12. Brenner BM, Anderson S. Glomerular function in diabetes mellitus. Adv Nephrol Necker Hosp 1990, 19:135–44.

13. Committee on Diabetic Twins JDS. Diabetes mellitus in twins: a cooperative study in Japan. Diabetes Res Clin Pr 1988, 5:271–80.

14. Knowler WC, Nelson RG, Pettitt DJ. Diabetes, Hypertension, and Kidney Disease in the Pima Indians. Kidney Hypertens. Diabetes Mellit., Springer; 1998, p. 141–50.

15. Mohan V, Vijayaprabha R, Rema M. Vascular complications in long-term South Indian NIDDM of over 25 years' duration. Diabetes Res Clin Pract 1996, 31:133–40.

16. Seaquist ER, Goetz FC, Rich S, Barbosa J. Familial clustering of diabetic kidney disease. N Engl J Med 1989, 320:1161–5.

17. Loe H. Periodontal Disease: The sixth complication of diabetes mellitus. Diabetes Care 1993, 16:329–34.

18. Gusberti FA, Syed SA, Bacon G, Grossman N, Loesche WJ. Puberty Gingivitis in Insulin-dependent Diabetic Children: I. Cross-sectional Observations. J Periodontol 1983, 54:714–20.

19. Safkan-Seppälä B, Ainamo J. Periodontal conditions in insulin-dependent diabetes mellitus. J Clin Periodontol 1992, 19:24–9.

20. Oliver RC, Tervonen T. Periodontitis and tooth loss: comparing diabetics with the general population. J Am Dent Assoc 1993, 124:71–6.

21. Deshpande K, Jain A, Sharma R, Prashar S, Jain

R. Diabetes and periodontitis. J Indian Soc Periodontol 2010, 14:207.

22. Galea H, Aganovic I, Aganovic M. The dental caries and periodontal disease experience of patients with early onset insulin dependent diabetes. Int Dent J 1986;36:219–24.

23. Harrison R, Bowen WH. Periodontal health, dental caries, and metabolic control in insulindependent diabetic children and adolescents. Pediatr Dent 1987, 9:283–6.

24. Ainamo J, Lahtinen A, Uitto V. Rapid periodontal destruction in adult humans with poorly controlled diabetes A report of 2 cases. J Clin Periodontol 1990, 17:22–8.

25. Cutler CW, Eke P, Arnold RR, Van Dyke TE. Defective neutrophil function in an insulin-dependent diabetes mellitus patient. A case report. J Periodontol 1991, 62:394–401.

26. Hayden P, Buckley LA. Diabetes mellitus and periodontal disease in an Irish population. J Periodontal Res 1989, 24:298–302.

27. Nichols C, Laster LL, Bodak-Gyovai LZ. Diabetes mellitus and periodontal disease. J Periodontol 1978, 49:85–8.

28. Rylander H, Ramberg P, Blohme G, Lindhe J. Prevalence of periodontal disease in young diabetics. J Clin Periodontol 1987, 14:38–43.

29. Sandholm L, Swanljung O, Rytomaa I, Kaprio EA, Maenpaa J. Periodontal status of Finnish adolescents with insulin-dependent diabetes mellitus. J Clin Periodontol 1989, 16:617–20.

30. Sandholm L, Swanljung O, Rytömaa I, Kaprio EA, Mäenpää J. Morphotypes of the subgingival microflora in diabetic adolescents in Finland. J Periodontol 1989, 60:526–8.

31. Sastrowijoto SH, Hillemans P, Steenbergen TJM, Abraham-Inpijn L, Graaff J. Periodontal condition and microbiology of healthy and diseased periodontal pockets in type 1 diabetes mellitus patients. J Clin Periodontol 1989, 16:316–22.

32. Thorstensson H, Kuylenstiema J, Hugoson A. Medical status and complications in relation to periodontal disease experience in insulin-dependent diabetics. J Clin Periodontol 1996, 23:194–202.

33. Collin H-L, Uusitupa M, Niskanen L, Koivisto A-M, Markkanen H, Meurman JH. Caries in patients with non-insulin-dependent diabetes mellitus. Oral Surgery, Oral Med Oral Pathol Oral Radiol Endodontology 1998, 85:680-5.

34. Ship JA. Diabetes and oral health: an overview. J Am Dent Assoc 2003, 134:4S-10S.

35. Senadheera MD, Guggenheim B, Spatafora GA, Huang Y-CC, Choi J, Hung DCI, et al. A VicRK signal transduction system in Streptococcus mutans affects gtfBCD, gbpB, and ftf expression, biofilm formation, and genetic competence development. J Bacteriol 2005, 187:4064–76.

36. Zhuang PL, Yu LX, Liao JK, Zhou Y, Lin HC. Relationship between the genetic polymorphisms of vicR and vicK Streptococcus mutans genes and early childhood caries in two-year-old children. BMC Oral Health 2018, 18:1–6.

37. Deng DM, Liu MJ, Ten Cate JM, Crielaard W. The VicRK system of Streptococcus mutans responds to oxidative stress. J Dent Res 2007, 86:606–10.

38. Salman MI, Rashied RM, Hamad HSH. Study of Liver function Tests in Diabetes Type-2 patients in Ramadi City, Iraq. Ann Trop Med Public Heal 2020, 23:231–824.

39. Lamster IB, Lalla E, Borgnakke WS, Taylor GW. The relationship between oral health and diabetes mellitus. J Am Dent Assoc 2008, 139:19S-24S.

40. Al-Ajlan AR. Lipid profile in relation to anthropometric measurements among college male students in Riyadh, Saudi Arabia: a cross-sectional study. Int J Biomed Sci IJBS 2011, 7:112.

41. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. Lancet 2005, 366:1809–20.

42. Soskolne WA, Klinger A. The relationship between periodontal diseases and diabetes: an overview. Ann Periodontol 2001;6:91–8.

43. Persson GR. Diabetes and Periodontal Disease: An Update for Health Care Providers. Diabetes Spectr 2011, 24:195–8.

44. Krishnan V, Thirunavukkarasu J. Assessment of knowledge of self blood glucose monitoring and extent of self titration of anti-diabetic drugs among diabetes mellitus patients–a cross sectional, community based study. J Clin Diagnostic Res JCDR 2016, 10:FC09.

45. Seethalakshmi C, Reddy RCJ, Asifa N, Prabhu S. Correlation of salivary pH, incidence of dental caries and periodontal status in diabetes mellitus patients: A cross-sectional study. J Clin Diagnostic Res JCDR 2016, 10:ZC12.

46. Al-Obaida MI, Al-Nakhli AKM, Arif IA, Faden A, Al-Otaibi S, Al-Eid B, et al. Molecular identification and diversity analysis of dental bacteria in diabetic and non-diabetic females from Saudi Arabia. Saudi J Biol Sci 2020, 27:358–62.

47. Latti B, Kalburge J, Birajdar S, Latti R. Evaluation of relationship between dental caries, diabetes mellitus and oral microbiota in diabetics. J Oral Maxillofac Pathol 2018, 22:282.

48. Hoshino T, Kawaguchi M, Shimizu N, Hoshino N, Ooshima T, Fujiwara T. PCR detection and identification of oral streptococci in saliva samples using gtf genes. Diagn Microbiol Infect Dis 2004, 48:195–9.

49. Saleh RO, Mohammed TK, Abdulmuhsen AM, Jwad MA. Variation of GTFD Gene from Streptococcus Mutans Local Isolate from Iraqi Patients. Indian J Public Health 2019, 10:189.

50. Almusawi MA, Gosadi I, Abidia R, Almasawi M, Alrashood ST, Ekhzaimy A, et al. Association between salivary factors and cariogenic bacteria in type-2 diabetes patients. J King Saud Univ 2020, 32:2617–21.

51. Rampini SK, Bloemberg G V, Keller PM, Büchler AC, Dollenmaier G, Speck RF, et al. Broad-range 16S rRNA gene polymerase chain reaction for diagnosis of culture-negative bacterial infections. Clin Infect Dis 2011, 53:1245–51.